

Dissertationes Forestales 67

Fen ecosystem carbon gas dynamics in changing
hydrological conditions

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Academic dissertation

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Fen ecosystem carbon gas dynamics in changing hydrological conditions

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ABSTRACT

Northern peatlands are thought to store one third of all soil carbon (C). Besides the C sink function, peatlands are one of the largest natural sources of methane (CH₄) to the atmosphere. Climate change may affect the C gas dynamics as well as the labile C pool. Because the peatland C sequestration and CH₄ emissions are governed by high water levels, changes in hydrology are seen as the driving factor in peatland ecosystem change.

This study aimed to quantify the carbon dioxide (CO₂) and CH₄ dynamics of a fen ecosystem at different spatial scales: plant community components scale, plant community scale and ecosystem scale, under hydrologically normal and water level drawdown conditions. C gas exchange was measured in two fens in southern Finland applying static chamber and eddy covariance techniques.

During hydrologically normal conditions, the ecosystem was a CO₂ sink and CH₄ source to the atmosphere. *Sphagnum* mosses and sedges were the most important contributors to the community photosynthesis. The presence of sedges had a major positive impact on CH₄ emissions while dwarf shrubs had a slightly attenuating impact. C fluxes varied considerably between the plant communities. Therefore, their proportions determined the ecosystem scale fluxes.

An experimental water level drawdown markedly reduced the photosynthesis and respiration of sedges and *Sphagnum* mosses and benefited shrubs. Consequently, changes were smaller at the ecosystem scale than at the plant group scale. The decrease in photosynthesis and the increase in respiration, mostly peat respiration, made the fen a smaller CO₂ sink. CH₄ fluxes were significantly lowered, close to zero. The impact of natural droughts was similar to, although more modest than, the impact of the experimental water level drawdown. The results are applicable to the short term impacts of the water level drawdown and to climatic conditions in which droughts become more frequent.

Keywords: peatland, carbon dioxide, methane, water level, climate change, functional groups

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LIST OF ORIGINAL ARTICLES

The thesis is based on the following articles which are referred to in the text by Roman numerals. The articles are reproduced with the kind permission from Springer Science + Business Media (Paper I) and Wiley-Blackwell Publishing Ltd. (Papers III, IV and V).

- I **Riutta, T.**, Laine, J. & Tuittila, E.-S. 2007. Sensitivity of CO₂ exchange of fen ecosystem components to water level variation. *Ecosystems* 10: 718-733. doi: 10.1007/s10021-007-9046-7.

- II **Riutta, T.**, Laine, J. & Tuittila, E.-S. The role of vegetation in methane dynamics in a boreal fen under pristine and water level drawdown conditions. Submitted manuscript.

- III **Riutta, T.**, Laine, J., Aurela, M., Rinne, J., Vesala, T., Laurila, T., Haapanala, S., Pihlatie, M. & Tuittila, E.-S. 2007. Spatial variation in plant community functions regulates carbon gas dynamics in a boreal fen ecosystem. *Tellus* 59B: 838-852. doi: 10.1111/j.1600-0889.2007.00302.x

- IV Aurela, M., **Riutta, T.**, Laurila, T., Tuovinen, J.-P., Vesala, T., Tuittila, E.-S., Rinne, J., Haapanala, S. & Laine, J. 2007. CO₂ exchange of a sedge fen in southern Finland — the impact of a drought period. *Tellus* 59B: 826-837. doi: 10.1111/j.1600-0889.2007.00309.x

- V Rinne, J., **Riutta, T.**, Pihlatie, M., Aurela, M., Haapanala, S., Tuovinen, J.-P., Tuittila, E.-S. & Vesala, T. 2007. Annual cycle of methane emission from a boreal fen measured by the eddy covariance technique. *Tellus* 59B: 449-457. doi: 10.1111/j.1600-0889.2007.00261.x

AUTHOR'S CONTRIBUTION

I & II The water level drawdown study was initiated in 2001. The author joined the research team in 2002. She participated in the planning and establishment of the extension of the study to include the ecosystem component aspect. She was responsible for the data collection during the years 2002 to 2004. She analyzed the data and wrote the papers upon which the co-authors commented.

III The author participated in the planning and establishment of the research site. She was responsible for the chamber flux and vegetation measurements. She participated in the set up and the maintenance of the eddy covariance flux measurements. She analyzed the chamber flux and vegetation data and wrote the paper upon which the co-authors commented.

IV & V The author participated in the planning and establishment of the research site, and in the set up and the maintenance of the eddy covariance flux measurements. She collected and analyzed the chamber flux (**IV**) and vegetation (**IV** and **V**) data. She participated in the interpretation of the results and helped write the papers.

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SUMMARY OF THE ARTICLES

I We studied the contribution of the fen plant community components, namely sedges, dwarf shrubs, *Sphagnum* mosses and the underlying peat, to the CO₂ fluxes in prevailing hydrological conditions and in water level drawdown conditions.

II We assessed the role of the fen plant community components, namely sedges, dwarf shrubs, *Sphagnum* mosses and the underlying peat in the CH₄ fluxes in prevailing hydrological conditions and in water level drawdown conditions.

III We described the plant community scale spatial variation in vegetation, and in CO₂ and CH₄ fluxes in a boreal fen. The spatial variation in the fluxes was determined with chamber measurements and the fluxes were upscaled to the ecosystem scale. The upscaled estimates were compared with the eddy covariance measurements. We assessed the sensitivity of the ecosystem fluxes to the spatial variation at the plant community scale.

IV We studied the CO₂ fluxes in a boreal fen ecosystem using the eddy covariance technique. We described the annual cycle, seasonal variation, and the response of the CO₂ fluxes to environmental, especially hydrometeorological, factors. Some plant community scale (chamber flux) data was included to strengthen the interpretation of the hydrometeorological responses.

V In this paper, we report the annual cycle and seasonal variation of CH₄ fluxes in a boreal fen, and the factors controlling the fluxes. To our best knowledge, this is the first continuous year-long time series of peatland methane fluxes measured by the eddy covariance technique.

1 INTRODUCTION

1.1 Background

Peatlands are ecosystems sustained by water table levels close to the surface where organic matter accumulates as peat. Northern peatlands cover approximately 4 million km², ca. 3% of the Earth's land area (Lappalainen 1996, Charman 2002). Of the total peatland area, the proportion of boreal and subarctic peatlands is approximately 85% (Joosten and Clarke 2002). In terms of land area, peatlands are a minor feature in the global landscape. However, the accumulated peat, the mass of which about half is carbon, makes them an important carbon reservoir. Up to one third of all terrestrial carbon, 250 to 400 Pg, is stored in northern peatlands (Gorham 1991, Turunen et al. 2002). This equals 30-50% of the carbon pool in the atmosphere (Denman et al. 2007). On the other hand, peatlands among other wetlands are the largest natural source of methane (CH₄) to the atmosphere (Matthews and Fung 1987, Mikaloff Fletcher et al. 2004). The high water levels and the consequent anoxic conditions that support carbon sequestration are also suitable for CH₄ production.

Human activities, such as fossil fuel burning and land use, have significantly increased the radiation absorbing gases in the atmosphere, most importantly carbon dioxide (CO₂), CH₄ and nitrous oxide (N₂O), leading to an enhanced greenhouse effect. Based on ice records, the current atmospheric CO₂ concentration (approximately 380 ppm) exceeds by far the natural range (180-300 ppm) during the last 650 000 years. The estimated total temperature increase from 1850–1899 to 2001–2005 is in the range of 0.6-1°C and the warming has been most rapid during the last 50 years. The warmth of the last half century is unusual in at least the previous 1,300 years. Most of the observed increase in global average temperatures since the mid-20th century is very likely due to the observed increase in anthropogenic greenhouse gas concentrations. Further warming and larger changes in the global climate system will occur if the greenhouse gas emissions continue at or above the current rates. (IPCC 2007)

Carbon accumulation in peatlands reduces the CO₂ burden of the atmosphere. Because of the CH₄ emissions, however, the impact of peatlands on the climate system is dual. In a time horizon of 100 years, a pulse emission of CH₄ has a 23 times larger radiative forcing impact than a pulse emission of the same mass of CO₂ (Ramaswamy et al. 2001). Therefore, if the annual CH₄ emissions and CO₂ sequestration of a peatland are considered as pulse events, the greenhouse warming potential (GWP) balance in many northern peatlands over a 100 year time scale is positive, i.e. the warming impact of the CH₄ emissions exceeds the cooling impact due to CO₂ sequestration (Whiting and Chanton 2001). However, peatlands have persistent, rather than pulse, CO₂ sequestration and CH₄ emission. If the radiative forcing impact is calculated for the entire Holocene, taking into account that CO₂ has a longer atmospheric lifetime than CH₄, the existence of peatlands have had a net cooling impact on the global climate, dominated by the persistent CO₂ sink function (Frolking and Roulet 2007).

Growing concern about global climate change has made the carbon cycle and carbon accumulation in peatlands a research priority. Biotic feedbacks from ecosystems on the climate system and the lability of carbon stocks in peatlands have been identified as some of the key uncertainties and research priorities by the Intergovernmental Panel on Climate Change (Fischlin et al. 2007). Northern peatlands have been resilient ecosystems, persisting

through Holocene climatic variation during which the mean annual temperature in the boreal and arctic regions has varied approximately 3°C above and 4°C below the current value (Andreev and Klimanov 2000, Antonsson and Seppä 2007). However, because climate change is projected to be most severe at the latitudes where northern peatlands are situated (Christensen et al. 2007), changes may be too rapid or too extreme for peatlands to maintain their stability.

1.2 Carbon cycle in peatlands

Phototrophic primary producers fix atmospheric CO₂ into organic compounds using radiative energy. Photosynthesis rates depend on photosynthetically active radiation (PAR), CO₂ concentration, water supply, temperature, and photosynthesizing plant biomass. Photosynthates (products of photosynthesis) are allocated to growth, respiration and storage, leached as root exudates or used by mycorrhizae. Compared to other terrestrial ecosystems, such as grasslands and forests, photosynthesis rates in peatlands are low (Frolking et al. 1998) Some 40 to 70% of the carbon fixed in photosynthesis is used in plant respiration (Gifford 2003, Litton et al. 2007). Consideration of plant respiration often includes the respiration of the rhizosphere microbes and mycorrhiza because their respiration rates are difficult to separate from other root functions (Chapin et al. 2006). The term ‘plant-derived respiration’ accounts for both the plant respiration and the heterotrophic rhizosphere respiration.

Dead plant litter and other soil organic matter, such as dissolved organic carbon, is decomposed by heterotrophic soil organisms (microbes and soil fauna), primarily under aerobic conditions. This process is called soil respiration or heterotrophic respiration. Soil respiration rates depend upon temperature, moisture and oxygen (O₂) availability, decomposer community, nutrient availability, and substrate availability and quality (Laiho 2006). Water level divides the peat layer into the oxic zone above and the anoxic water saturated zone below the water level. O₂ diffusion to the water saturated zone is very slow because the diffusion of gases is approximately 10⁴ times slower in water than in air. The aerobic decomposition of the organic matter to CO₂ in the oxic zone is fast compared to the anaerobic decomposition of organic matter to CH₄ in the anoxic zone.

If there is an imbalance between the photosynthesis and the plant and soil respiration the incompletely decomposed organic matter will accumulate as peat. The slow decomposition rate is the main reason for the carbon accumulation in peatlands (Clymo 1983). The decomposition of organic matter in peatlands is slow because of the small volume of the oxic layer, low soil temperatures during large part of the year and the poor decomposability of the plant material, particularly *Sphagnum* moss tissue. Over millennial time scales, the persistent carbon sink function has created substantial carbon reservoirs as peatlands have expanded and grown in height. However, most of the assimilated CO₂ is released back into the atmosphere via autotrophic or heterotrophic respiration and only a small fraction of the gross primary production, up to 15% annually (Clymo 1984, Gorham 1991, Francez and Vasander 1995), is sequestered in the peat. Therefore, even small changes in the CO₂ uptake or release can cause considerable variation in the annual CO₂ balances, which can be small, variable, and dependent on the weather conditions (e.g. Lafleur et al. 2003, Aurela et al. 2004, Roulet et al. 2007). As a result, a peatland may readily switch from a CO₂ sink to a CO₂ source between cold and wet and hot and dry years (Alm et al. 1999b).

Because of the high water levels and relatively high rates of carbon supply, conditions in peat are often anoxic. Anoxic conditions and the availability of suitable carbon sources are prerequisites for CH₄ production. CH₄ is a metabolic end product of strictly anaerobic microbes that belong to the domain Archaea. In peatlands, CH₄ is formed either from acetate dissimilation (acetate pathway) or bicarbonate reduction (hydrogen pathway) (Zinder 1993) after a complex organic matter decomposition chain. The acetate pathway dominates in vegetated sites where fresh organic matter is available, while the hydrogen pathway dominates or co-dominates in unvegetated sites, outside of the growing season or in bogs (Bellisario et al. 1999, Popp et al. 1999, Chasar et al. 2000, Keller and Bridgman 2007). Methanogenesis may be suppressed if the concentrations of alternative electron acceptors such as nitrate (NO₃⁻), sulfate (SO₄⁻²), ferric iron (Fe(III)), manganese (Mn(II)) or humic acids are high (Lovley and Klung 1986, Lovley et al. 1996).

CH₄ is released from the anaerobic peat layers to the atmosphere via diffusion through the peat matrix, passage through plants or ebullition. Diffusion through the peat matrix, especially through the water saturated layer is slow because of the slow diffusion of gases in the liquid phase. Compared to the diffusion through peat matrix, passage through the plant tissue is fast. CH₄ molecules move through the vascular plant tissue by passive diffusion along the concentration gradient (Schimel 1995, Shannon et al. 1996) or by an active gas transport system, such as pressure induced convection (Dacey and Klug 1979, Brix et al. 1992). Plant-mediated transport dominates in the presence of vascular plants (Sebacher et al. 1985, Whiting and Chanton 1992, Schimel 1995) while ebullition dominates in the absence of vascular plants (van der Nat et al. 1998). Bubbles may be formed if gases become supersaturated in the pore water. The mass of stored CH₄ as gas bubbles can be as much as three times the mass of dissolved CH₄, depending on the time of year (Fechner-Levy and Hemond 1996). Changes in air pressure and fluctuations in water level may trigger a degassing of the bubble pool, resulting in large episodic fluxes that may contribute significantly to the annual or seasonal fluxes also in vegetated sites (Moore et al. 1990, Glaser et al. 2004, Tokida et al. 2005).

A considerable part, up to 100% (Le Mer and Roger 2001, Pearce and Clymo 2001, Whalen 2005), of the CH₄ diffusing through the upper, aerobic peat layer can be oxidized to CO₂ by methanotrophic and methylotrophic bacteria before reaching the atmosphere. The net CH₄ flux is the balance between the CH₄ production, CH₄ consumption, transport rate and storage.

Peatland CH₄ fluxes are regulated by temperature, water level, substrate availability and quality, and variations in weather conditions. Water level regulates the volume ratio between the aerobic and anaerobic zones and, consequently, the extent of the CH₄ production and oxidation zones. Therefore, in most cases there is a positive correlation between the water level and CH₄ fluxes (Clymo and Reddaway 1971, Moore and Roulet 1993, Laine et al. 2007b). The activity of methanogenic and methanotrophic microbes is temperature dependent. CH₄ production has a stronger temperature response than CH₄ oxidation (Dunfield et al. 1993). Therefore, CH₄ fluxes generally increase with increasing temperature (Moore and Dalva 1993, Daulat and Clymo 1998, Bellisario et al. 1999, Christensen et al. 2003) but the response can also be negative (MacDonald et al. 1998). New, recently fixed carbon is a more easily decomposable substrate for the anaerobic decomposition chain compared to the old, recalcitrant peat. Therefore, CH₄ fluxes should correlate positively with net ecosystem CO₂ exchange (NEE) (Whiting and Chanton 1993, Bellisario et al. 1999, Christensen et al. 2000). However, the controlling factors of the CH₄ flux are not independent but there are complex interactions. For example, the temperature

effect can also be enhanced or constrained by substrate availability (Whiting and Chanton 1993, Bergman et al. 1998) and substrate type (Svensson 1984), and the importance of the plant-mediated CH₄ flux depends on the water table level (Waddington et al. 1996): the deeper the water level, the fewer roots and photosynthates reach the anaerobic layer of the peat. Owing to these interactions and the individual responses of CH₄ production, oxidation and transport to the controlling factors, identifying the responses of the net flux is difficult and different studies show different results, depending on the climatic conditions and site characteristics.

In comparison to CO₂ uptake and efflux, CH₄ fluxes are small. However, they can be an important component of the peatland carbon balance (Table 1). Another important component is dissolved organic carbon. Loss of carbon in that form can be up to 23 to 37% of the annual carbon balance (Waddington and Roulet 2000, Roulet et al. 2007). The estimates for the long term average carbon accumulation rates in northern peatlands range from 17 to 29 g C m⁻² in different parts of the boreal region (Clymo et al. 1998, Gorham 1991, Korhola et al. 1995, Laine et al. 1996, Turunen et al. 2002). However, these rates may not reflect current or future situations. Recent studies on pristine peatlands have shown that annual CO₂ balances and CH₄ efflux rates are highly variable, both in space (within and between sites) and in time (Table 1).

1.3 Role of vegetation in peatland carbon cycle

Peatlands can be divided into fens that receive water and nutrient inputs from the surrounding mineral soils and bogs that receive water and nutrients solely from the atmospheric inputs. The hydrological dynamics of the system and the amount of incoming nutrients are the main factors controlling species occurrence in peatlands (Wheeler and Proctor 2000, Økland et al. 2001). The anoxic rooting environment impedes the growth of plants which have not adapted to such conditions, including most tree species (Jeglum 1974, Macdonald and Yin 1999). Plant communities in Eurasian fens typically consist of sedges, dwarf shrubs and *Sphagnum* mosses (Botch and Masing 1983, Ruuhijärvi 1983, Sjörs 1983). Dividing the species into functional plant groups has provided a useful simplification when studying ecosystem change because group members are likely to have similar responses to environmental factors (Aguiar et al. 1996, Chapin et al. 1996).

Peatland plants differ in their photosynthetic capacity (Bubier et al. 2003b), respiration (Bubier et al. 2003b), quality of produced litter (Szumigalski and Bayley 1996), and role in CH₄ dynamics (Shannon et al. 1996, Joabsson et al. 1999). Therefore, the plant composition of a peatland ecosystem influences the autotrophic CO₂ exchange rates as well as the soil processes. Because plant groups respond differently to environmental factors, a change in environmental conditions alters the contribution of the different species to the community CO₂ and CH₄ exchange (Karlsson 1989, Lovelock and Feller 2003, Strack et al. 2006b) or to the community biomass accumulation and allocation (Parsons et al. 1994, Wijesinghe et al. 2005). Eventually internal (ecological succession) and external (weather and hydrological conditions) forcings modify not only the carbon dynamics but also the autotrophic and heterotrophic communities, which in turn feeds back on the carbon cycle.

Table 1. Observed carbon fluxes in some undrained sedge-dominated fens, measured using eddy covariance (EC) or chamber method. Unit of the fluxes is g C m^{-2} , and the range of the fluxes is shown. "Spatial" is the range of fluxes between vegetation types within one study year and "Temporal" is the range of spatially averaged fluxes between study years. The number of vegetation types / study years is given in parenthesis. Positive NEE indicates a CO_2 sink and negative NEE a CO_2 source to the atmosphere.

Site	Method	Latitude; Longitude	MAT ^a	Annual NEE	
		N; E	°C	Spatial	Temporal
Zackenbergl, Greenland	EC	74; 20	-19.5		64 (1)
Barrow, Alaska, USA	Chambers	70; 156	-12.6		
Churchill, Manitoba, Canada	EC	58; 94	-7.2		
Schefferville, Québec, Canada	Chambers	54; 66	-4.9		
Lek Vorkuta, Komi Republic, Russia	Chambers	67; 63	-6.0	-3.0 - 43 (5)	
James Bay Coast, Québec, Canada	Chambers	54; 78	-3.1		
Kaamanen, Finland	EC	69; 27	-1.1		4.1 - 53 (6)
Degerö, Sweden	EC	61; 19	1.2		48 - 61 (3)
Salmisuo, Finland	Chambers	62; 30	1.9	53 - 105 (4)	
Lakkasuo, Finland	Chambers	61; 24	3.4		
Siikaneva, Finland	Chambers, EC	61; 24	3.4		51 - 60 (2)
Sallie's Fen, New Hampshire, USA	Chambers	43; 71	8.1		
Big Cassandra, Michigan, USA (BC4)	Chambers	42; 84	8.3		

^a Mean annual temperature

^b Range of individual sample plots instead of vegetation types

Growing season NEE		Annual CH ₄ emission		Growing season CH ₄ emission		Reference
Spatial	Temporal	Spatial	Temporal	Spatial	Temporal	
	96 (1)					Soegaard and Nordstroem (1999)
	-14 - 4 (5)					Oechel et al. (1995)
	-76 - 235 (5)					Griffis et al. (2001)
		0.98 - 7.4 (5)				Moore and Knowles (1990)
1.0 - 49 (5)		1.2 - 12 (5)		0.1 - 8.2 (5)		Heikkinen et al. (2002)
				0.47 - 13 (5)		Pelletier et al. (2007)
			5.5 (1)			Aurela et al. (2004); Hargreaves et al. (2001)
						Sagerfors et al. (2008)
108 - 160 (4)		13 - 36 (4)		11 - 30 (4)		Alm et al. (1997)
	32 - 130 (4)				2.6 - 6.8 (4)	Papers I and II
21-193 (5)			9.4 (1)	5.4 -18 (5)		Papers III, IV and V
				16 - 62 (11) ^b	17 - 39 (5)	Treat et al. (2007)
			3.1 - 36 (3)			Shannon and White (1994)

Sedges, family *Cyperaceae* (e.g. genera *Carex*, *Eriophorum*, and *Trichophorum*), are perennial grasses that reproduce mainly vegetatively (Bernard 1976, Bedford et al. 1988). Sedges produce new shoots throughout the growing season while old shoots die (Bernard 1976, Bedford et al. 1988). The shoots produced in late autumn overwinter (Bernard et al. 1988), which ensures high photosynthetic capacity during the entire growing season. Sedges are deep-rooting and allocate approximately 90% of their total biomass to the roots (Sjörs 1991, Saarinen 1996). The root biomass is mainly located in the top 25 to 30 cm layer (Sjörs 1991, Saarinen 1996), but some *Carex rostrata* Stokes roots have been found as deep as 230 cm (Saarinen 1996). Because of the deep roots, new organic matter is simultaneously injected at different depths of the peat profile in sedge-dominated sites, not just on the surface layer. Sedges have aerenchymatous tissue, an adaptation to wet conditions, for transporting molecular O₂ from the above ground parts to the roots (Fagerstedt 1992, Moog and Bruggemann 1998, Visser et al. 2000). Consequently, sedges can tolerate anaerobic, water-logged conditions and are strong competitors in such habitats. Sedges are most abundant in minerotrophic peatlands where the water level is close to the surface (Laine and Vanha-Majamaa 1992).

Sedges are particularly important in peatland CH₄ dynamics; the deep roots produce fresh substrate, with relatively little structural carbohydrates compared to woody species, for the methanogenic consortia in the anoxic peat layer and the aerenchymatous tissue serves as a conduit for CH₄ molecules from the soil to the atmosphere, bypassing the oxic surface layer. Plant mediated transport accounts for a major fraction (75-97%) of the total CH₄ emissions in sedge dominated sites (Whiting and Chanton 1992, Schimel 1995, Kelker and Chanton 1997). On the other hand, O₂ transport through the aerenchyma to the rhizosphere inhibits CH₄ production (Whalen and Reeburgh 2000) and stimulates CH₄ oxidation (King 1994, Popp et al. 2000). All in all, the net effect of the presence of aerenchymatous species is to increase CH₄ fluxes in most cases (Waddington et al. 1996, Frenzel and Rudolph 1998, King et al. 1998, Bellisario et al. 1999, Greenup et al. 2000, Rinnan et al. 2003).

Dwarf shrubs are a diverse group consisting of both deciduous and evergreen species. Shrub photosynthesis is less sensitive to environmental conditions (Bubier et al. 2003a) and the shoot growth depends more on the age of the individual ramet (stems that had emerged above the moss layer which might not be genetic individuals) than on climatic variables (Shevtsova et al. 1995). The fundamental difference between these woody plants and herbaceous sedges is that the above ground growth of shrubs is cumulative whereas sedges have to rebuild their aboveground parts each growing season. Some CH₄ can be emitted through woody species though at considerably lower rates than through aerenchymatous species (Shannon et al. 1996, Garnet et al. 2005). Recently, it was suggested that terrestrial plants would produce and emit significant amounts of CH₄ (Keppler et al. 2006). The findings were then questioned (Dueck et al. 2006), but later an independent data set demonstrated CH₄ emissions from shrubs (Wang et al. 2008). A probable source of CH₄ is methoxyl groups of pectin split by UV radiation (Keppler et al. 2008).

Sphagnum mosses are the dominant moss genus in boreal peatlands. *Sphagnum* can tolerate very wet, acidic and nutrient poor conditions (Clymo and Hayward 1982) and photosynthesize efficiently at low temperatures (Harley et al. 1989), as long as the moss surface is not frozen (Malmer et al. 2003). Like other bryophytes, they are poikilohydric plants that lack internal water conducting structures, which makes their physiological processes highly sensitive to water availability. *Sphagnum* mosses have a species specific optimal water content for photosynthesis and a narrow tolerance to water content variation

(Silvola and Aaltonen 1984, Rydin and McDonald 1985b, Gaberscik and Martincic 1987, Schipperges and Rydin 1998).

Different litter materials may have markedly different decomposition rates which influences the peatland carbon accumulation rates (Laiho 2006). Of the three functional groups described above, the litter produced by *Sphagna* is the most decomposition resistant, the litter produced by sedges is the least decomposition resistant and the litter produced by shrubs is intermediate (Szumigalski and Bayley 1996).

Typically, a fen ecosystem comprises a continuum of microforms along the water level gradient, from dry hummocks through lawns to wet hollows. The microforms have distinct plant community composition. The relative abundance of sedges and shrubs changes from the sedge dominance in the hollows to the shrub dominance in the hummocks. *Sphagnum* mosses are present throughout the continuum but the species composition varies. *Sphagnum* mosses hold species-specific niches in the hummock-hollow gradient (Vitt and Slack 1984, Rydin 1985) having morphological adaptation to different moisture condition (Clymo and Hayward 1982). Spatial variation in C gas fluxes between communities is very high (Table 1, Svensson and Rosswall 1984, Moore and Knowles 1990, Waddington and Roulet 2000, Laine et al. 2007a, Laine et al. 2007b). In patterned ecosystems, the proportions of the communities determine the ecosystem fluxes (Laine et al. 2006).

1.4 Climate change impact on peatland ecosystems

Although most climate models predict an increase in global temperatures and precipitation, the changes are not evenly distributed among the different regions of the globe (Christensen et al. 2007). There is higher uncertainty in the regional projected changes in precipitation than in temperature. Some climate models predict a water level drawdown in northern peatlands. Using the scenario of a doubled CO₂ concentration, leading to a 3°C increase in the June, July and August monthly mean temperatures and a 1 mm increase in daily precipitation (Mitchell 1989) Roulet et al. (1992) estimated a water level drawdown of 14 to 22 cm in northern peatlands, due to the increased evapotranspiration which more than compensates the effect of the increased precipitation. Water level drawdown, the indirect effect of the global warming, is the most important effect of the climate change on peatlands, overshadowing the direct temperature effect, the effect of the elevated atmospheric CO₂ concentration, and the effect of the prolonged growing season (Gorham 1991, Gitay et al. 2001, Moore 2002). The water level effect dominates, because water level regulates the plant community composition and the volume ratio of the oxic and anoxic zones and therefore, the relative rates of aerobic and anaerobic decomposition.

Studies on the short term impact of water level drawdown, caused either by natural variability in weather conditions or by treatment, have demonstrated decrease in photosynthesis (Alm et al. 1999b, Bubier et al. 2003a), increase in respiration (Alm et al. 1999b, Bubier et al. 2003a, Strack et al. 2006a), decreased CO₂ sink function (Alm et al. 1999b, Bubier et al. 2003a, Lafleur et al. 2003), and decrease in CH₄ emissions (Strack and Waddington 2007). However, the responses are not unidirectional but depend on the initial water level conditions and on the vegetation composition (Bubier et al. 2003a, Bubier et al. 2003b, Strack et al. 2004, Strack et al. 2006a).

Over the long term, permanently drier conditions will lead to a vegetation change in northern peatlands. A moderate water level drawdown makes them suitable to a larger number of species (Visser et al. 2000, Weltzin et al. 2003) and the plants that are adapted to

high water levels lose their competitive advantage. Studies on the impact of forestry drainage have shown that following the water level drawdown of 20-50 cm, a sedge and *Sphagnum* dominated community will turn into a pine and forest moss dominated community in a few decades (Laine et al. 1995, Laiho et al. 2003).

1.5 Aims of the study

The general aim of the study was to quantify the CO₂ and CH₄ dynamics of fen plant community components (sedges, dwarf shrubs, *Sphagnum* mosses and the underlying peat), a continuum of fen plant communities, and the fen ecosystem as a whole, under hydrologically normal and water level drawdown conditions. More specifically:

The aim of the plant community component scale studies (Papers **I** and **II**) was to assess the contribution of the plant community components to the CO₂ and CH₄ fluxes in different water level conditions and the sensitivity and response of the components to water level variation.

The aim of the plant community scale study (Paper **III**) was to define the plant communities of the fen, to quantify the CO₂ and CH₄ exchange of the communities and to examine how the spatial variation at the plant community scale is reflected in the carbon gas dynamics at the ecosystem scale.

The aim of the ecosystem scale studies (Papers **IV** and **V**) was to quantify the annual CO₂ and CH₄ exchange of the fen, temporal variation in the C gas exchange, and the response to environmental, especially hydrometeorological, factors.

2 MATERIALS AND METHODS

2.1 Study sites

2.1.1 Study region

The study was carried out at two sites in southern Finland, 60 km from the city of Tampere. The sites are situated approximately 6 km from the Hyytiälä Forestry Field station and 8 km from one another. Annual precipitation in the region totals 710 mm, of which about a third falls as snow. The average temperatures for January and July are - 8.9 and 15.3 °C, respectively, and the average cumulative temperature sum ($\geq +5^{\circ}\text{C}$) is 1160 degree days (Juupajoki-Hyytiälä weather station, Drebs et al. 2002). The region lies in southern boreal vegetation zone, close to the border of the middle boreal zone (Ahti et al. 1968).

2.1.2 Lakkasuo study site

Lakkasuo is an eccentric raised bog complex with extensive minerotrophic margins. This study was carried out at an oligotrophic, treeless fen site (61 47' N; 24 18' E). In the field layer, sedges (*Carex lasiocarpa* Ehrh., *Eriophorum vaginatum* L.) are the dominant plant group. In addition, dwarf shrubs (*Betula nana* L., *Andromeda polifolia* L. and *Vaccinium oxycoccos* L.) make up a considerable proportion of the field layer. The moss layer is a

continuous peat moss carpet dominated by *Sphagnum papillosum* Lindb., *S. fallax* (Klinggr.) Klinggr. and *S. flexuosum* Dozy & Molk.

In Lakkasuo, we studied the C gas exchange at the plant community component scale (Papers **I** and **II**) in prevailing and water level drawdown conditions, using a closed chamber method. The study site was divided into two subsites, namely the control and the water level drawdown subsites. The water level drawdown subsite was surrounded with a shallow ditch that lowered the water level by 10 to 25 cm. The measurements were carried out during four growing seasons, 2001 to 2004. The first year was a calibration season when both subsites were measured prior to the water level drawdown treatment. Water level treatment was implemented in April 2002.

The contribution of the plant community components was studied by means of vegetation removal treatments. In both control and water level drawdown subsites, there were four types of permanent gas exchange sample plots consisting of:

- peat, *Sphagnum* mosses, sedges, and shrubs (intact vegetation)
- peat, *Sphagnum* mosses, and sedges (shrubs removed)
- peat and *Sphagnum* mosses (shrubs and sedges removed)
- peat (all vegetation removed).

The contribution of each plant community component to the CO₂ and CH₄ fluxes was assessed by comparing the fluxes in the different plot types. In 2001, only the plots with intact vegetation were established. The removal treatment plots were added in 2002. Above-ground parts of the vascular plants, and the top two cm of the moss layer was removed. Emerging re-growth was clipped off once a week, if necessary. Progressively less clipping was needed over the course of the study, hardly any during the third removal treatment year (2004). After clipping, soil respiration can decrease significantly and reach a new steady state as fast as in two days (Bremer and et al. 1998, Wan and Luo 2003). Therefore, CO₂ flux data from all removal treatment years could be used in the analyses (Paper **I**). The CH₄ flux data, on the other hand, showed a highly irregular pattern, indicating considerable disturbance, during the first two years of the treatment (Paper **II**). Therefore, only the data from the growing season 2004 was used.

2.1.3 Siikaneva study site

Siikaneva is the second largest undrained peatland complex in the southern part of the country. It is comprised of raised bogs, southern aapa fens and upland forest patches. Peatlands make up 80% of the total area. The study site (61°49' N, 24°11' E) is situated in an open oligotrophic fen part of the complex. We studied the C gas exchange at the ecosystem scale using an eddy covariance (EC) technique (Papers **IV** and **V**) and at the plant community scale using a chamber technique (Paper **III**). The site was suitable for an ecosystem scale study, as it offers a large, open and relatively homogeneous fetch, and for the plant community scale study because it comprises a continuum of communities. Although the microforms in Siikaneva site are not very pronounced, the site has a gradient from dry hummock to intermediate lawns and wet hollows. Lawns are the dominant microform, covering some 75% of the site and can be further classified into different types based on their species composition.

Dominant vascular plant species are sedges *Eriophorum vaginatum*, *Carex rostrata* Stokes, and *C. limosa* L., the shrubs *Andromeda polifolia* L. *Betula nana* L., and the herb *Rubus chamaemorus* L. *Sphagnum* mosses form a continuous carpet. Dominant species are

Sphagnum balticum (Russow) Russow ex C.E.O. Jensen and *S. papillosum* in lawns, *S. magellanicum* Brid. in hummocks, and *S. majus* (Russow) C.E.O. Jensen in hollows.

In Siikaneva, we established 24 permanent gas exchange sample plots that covered the variation in vegetation and water level. Vegetation was removed from six plots to examine the contribution of the soil processes to the fluxes. Infrastructure was built and the sample plots were established in the autumn 2003. Measurements were started in the spring 2004. During 2004, we measured CO₂ and CH₄ fluxes at the plant community scale and CO₂ fluxes at the ecosystem scale. In February 2005, we began to measure also CH₄ fluxes at the ecosystem scale.

2.2 Measurement methods

2.2.1 Fluxes measured using closed chamber and snowpack diffusion methods

Closed chambers are containers that, when placed on the measurement spot, isolate a piece of the ecosystem from the surrounding atmosphere. The measurement of the net flux is obtained as the rate of change in gas concentration during a known period of time. The source area of the flux is quite well defined, especially if permanent measurement collars are inserted into the soil. The chamber method allows the quantification of spatial variation in the fluxes, the quantification of the properties of the source area, and the examination of the relationship of the fluxes with those properties. In addition, experimental manipulations are relatively easy to carry out. CO₂ fluxes can be partitioned into gross photosynthesis and respiration by conducting successive measurements in light and dark. A serious limitation of the chamber method is the disturbance of the measurement collar and the chamber to the system. Manual chamber measurements are labor intensive and thus lead to poor temporal resolution. Therefore, episodic fluxes are unlikely to be captured reliably.

In this study, plant community and plant community component scale fluxes were measured by the manual chamber technique (Papers **I**, **II** and **III**). Permanent rectangular (56×56 cm) measurement collars were inserted to a depth of 30 cm into each gas exchange sample plot to isolate the majority of the roots. On the top of the collar there was water groove that allowed chamber placement and air-tight sealing of the measurement system. Chamber measurements were carried out at weekly or biweekly intervals during the snow-free season.

Instantaneous CO₂ flux was measured with a temperature controlled transparent plastic chamber of 60×60×30 cm (NSNF-1 in Pumpanen et al. 2004, Alm et al. 2007). CO₂ concentration in the chamber was monitored with a portable infrared CO₂ analyzer (EGM-2, EGM-3, EGM-4, PP Systems, UK) for 90 to 180 seconds. CO₂ concentration, PAR, and temperature inside the chamber were recorded at 15 second intervals. To determine the dependence of the CO₂ exchange rate on PAR, a series of measurements were performed at each plot, first in full light, then under shades of varying thickness and lastly in the dark. The chamber was lifted from the collar between the measurements to restore the ambient conditions.

An opaque aluminum chamber of 60×60×30 cm was used in the CH₄ flux measurements, (Crill et al. 1988). A 40 ml air sample was drawn to a polypropylene syringe at 5, 15, 25 and 35 minutes after closure. The samples were analyzed with a gas chromatograph (HP-5710A and HP-5890A) equipped with a flame ionization detector (GC-FID) within 36 hours. The performance of the instrument was evaluated by analyzing

calibration gas samples (CH₄ concentrations of 1.84 and 10.6 ppm). The precision of the analysis was 0.16%, determined as coefficient of variation of replicate calibration gas samples.

The emphasis of the plant community and plant community component scale studies was on the growing season fluxes. At the Siikaneva site, the spatial variation in the fluxes during the snow-covered season was assessed by occasional winter measurements. Chambers were used when the snow depth was < 20 cm, otherwise a snowpack diffusion method (Alm et al. 1999a) was applied. The chamber method was the same as the CH₄ measurement method during the snow-free season. In the snowpack diffusion method, gas samples from the top of the snow pack and from the moss surface were drawn into syringes using a metal pipe of 1-mm diameter. At each sampling point, the porosity of the snow was determined by weighting a volumetric snow sample throughout the depth of the snow layer and calculated using the density of pure ice (0.92 g cm⁻³). CH₄ concentrations in the samples were analyzed with the GC-FID and CO₂ concentrations with EGM-4 in the laboratory.

Flux rates in the chamber measurements were calculated as the linear rate of change in CO₂ or CH₄ concentration inside the chamber headspace. CO₂ flux measurements in the dark represented total (plant-derived and soil) respiration (R). An estimate for photosynthesis (P_G) was calculated by subtracting the CO₂ exchange rate in the light conditions from the exchange rate in the subsequent dark measurement. The flux in the snowpack diffusion method was calculated from the concentration difference between the top and the bottom layer as a function of the layer depth, snow porosity and snow temperature by applying Fick's first law of diffusion through porous media and using the diffusion coefficients of 0.139 cm² s⁻¹ and 0.22 cm² s⁻¹ for CO₂ and CH₄, respectively (Sommerfeld et al. 1993). The snow pack at each sampling point was assumed homogeneous, that is, density changes or ice lenses were not monitored.

2.2.2 Fluxes measured using eddy covariance

The ecosystem scale measurements were conducted with an eddy covariance (EC) method in Siikaneva site (Papers IV and V). The EC technique is a continuous micrometeorological flux measurement method. The flux is determined as the covariance of the correlation between the vertical wind velocity and the scalar of interest, such as temperature, water vapor, CO₂, or CH₄ concentration. Both the vertical wind velocity and the scalar are measured at similar rates with fast-response sensors typically at the rate of 10 Hz. The flux is obtained using the following general formula (cf. Baldocchi 2003):

$$F = \overline{\rho_a w' c'} \quad (1)$$

In the Equation 1, F is the flux, ρ_a is the air density, w is the vertical wind velocity and c is the mixing ratio of the scalar (e.g. CO₂). Overbars denote the mean of discrete time averaging and primes denote the residuals from the mean ($w' = w - \bar{w}$ and $c' = c - \bar{c}$). The fluxes in our studies are averaged over 30-minute periods.

Measurements are conducted above the vegetation canopy and the measured fluxes represent an average exchange rate from an area upwind from the measurement point. Therefore the flux estimates reflect the processes at the ecosystem scale. The source area, or the flux footprint, varies continuously with atmospheric stability and wind strength and direction. The change in the footprint does not affect the flux estimates if the spatial

variation in the ecosystems is small relative to the footprint. If the horizontal length scale of the heterogeneities is small enough the heterogeneities are smeared by turbulence below the measurement height. In heterogeneous ecosystems, however, the analysis of the source area and knowledge of the degree of spatial variation are necessary for the interpretation of the results (Schmid 2002, Laine et al. 2006)

EC has many advantages: it offers good temporal resolution and the gas exchange estimates can be calculated for different time scales, from half hours to years. The technique is a direct measurement of the flux at the ecosystem scale and the measurement system does not cause disturbance to the ecosystem. The limitations of the method include the assumption of the composition of the source area and often a poor night time data coverage due to the lack of turbulence.

The EC instrumentation in this study included a USA-1 (METEK, Germany) three-axis sonic anemometer/thermometer, a LI-7000 (Li-Cor Inc., USA) closed path analyzer for CO₂/H₂O and a tunable diode laser absorption spectrometer (TDL, TGA-100, Campbell Scientific Inc., USA) for CH₄. The measurement frequency was 10 Hz. The flux tower (measurement height 3.0 m) was located in a position where the open peatland area extends 200–400 m in all directions (see aerial photograph in Paper III). The chosen measurement height was low enough to keep the flux footprint within the open peatland under most conditions and high enough to spatially average the fluxes originating from individual elements composing the surface.

2.2.3 *Auxiliary measurements: meteorology and vegetation monitoring*

In order to relate the C gas fluxes to prevailing environmental conditions, meteorological parameters (air and peat temperatures, photosynthetically active radiation, precipitation, relative humidity, and water level) were measured continuously at weather stations in both study sites. In addition, water level, and air and peat temperatures in each sample plot were measured simultaneously with the chamber flux measurements.

The amount of vegetation, the species composition and the relative abundance of the species varies both spatially, between locations, and temporally; interannually and over the course of the growing season. Because vegetation is a fundamental controller of the C gas exchange it was important to monitor this variation.

In the permanent gas exchange sample plots, vegetation was monitored monthly during the snow-free period. We employed the method by Wilson et al. (2007) to obtain the vascular green leaf area in each sample plot. In short, the number of the green leaves of each species was calculated in every plot. The average size of the leaves was determined based on the leaf samples outside the plots. To these monthly observations, we fitted a unimodal curve to estimate the continuous development of the species-specific green area in each sample plot. The green area of mosses was determined as a percent cover of moss capitula once during each growing season.

In Siikaneva site, where the ecosystem scale fluxes were of interest, we also conducted a vegetation inventory (Paper III). This was a systematic inventory in a 30 m grid that extended to 200 m radius from the EC measurement mast. The inventory data allowed for the identification of the vegetation communities and the quantification of the proportions and average green areas of the communities in different parts of the site.

2.3 Data analysis

2.3.1 CO_2 fluxes measured using chamber method

The main method for analyzing the CO_2 flux data measured by chambers (Papers **I** and **III**) was the use of nonlinear regression models. The models served two purposes: 1) the reconstruction of the continuous time series of the CO_2 fluxes, and 2) the analysis of the response of the CO_2 dynamics to different biotic and abiotic factors.

Because photosynthesis reacts almost immediately to the amount of incoming light, which changes on a time scale of seconds, the rate of photosynthesis can vary very rapidly. Respiration varies over hourly time scales, responding to the variation in temperature and, with a time lag, photosynthesis (Kuzyakov and Cheng 2001, Tang et al. 2005). Therefore, the chamber flux measurements in the field, where environmental conditions cannot be standardized, reflect only a momentary CO_2 exchange rate. In this study, the measurements were conducted only weekly or biweekly and cannot be used directly to draw conclusions on the seasonal CO_2 exchange or on the differences between the sample plots. This problem can be overcome by modeling the response of the CO_2 flux using the environmental conditions during the measurements. With the models and continuous time series of the explanatory variables, it is possible to calculate a CO_2 flux rate in any given time. The models also reveal the potential differences in the responses among sample plots, vegetation communities or plant groups.

A semi-empirical modeling approach was adapted from Tuittila et al. (2004). The forms of the response functions have theoretical premises. The parameter values were derived empirically from the data using non-linear regression, but before a model was accepted, we checked that the parameter values were ecologically reasonable. We used PAR, the amount of green area, temperature, and water level as explanatory variable in photosynthesis models and the amount of green area, temperature, and water level as explanatory variables in respiration models. The default versions of the models are given in Eq. 2 (P_G) and Eq. 3 (R). However, not all responses could be identified in every case.

$$P_G = P_{max} \underbrace{\frac{PAR}{k + PAR}}_I \underbrace{\exp\left[-0.5\left(\frac{T - T_{opt}}{T_{tol}}\right)^2\right]}_{II} \underbrace{\exp\left[-0.5\left(\frac{WL - WL_{opt}}{WL_{tol}}\right)^2\right]}_{III} \times \underbrace{[1 - \exp(-\alpha GA)]}_{IV} \quad (2)$$

The Eq. (2) has the following explanatory variables:

PAR	photosynthetically active radiation ($\mu\text{mol m}^2 \text{s}^{-1}$)
T	air temperature inside the measurement chamber (C)
WL	water level relative to the moss surface (cm)
GA	green area ($\text{m}^2 \text{m}^{-2}$)

And the following parameters:

- P_{max} potential maximum rate of gross photosynthesis when all factors (PAR, T, WL, GA) are non-limiting
 k level of PAR at which P_G reaches 50% of the potential maximum rate
 T_{opt} optimum temperature for photosynthesis
 T_{tol} temperature tolerance, i.e. deviation from the optimum T at which P_G is 61% of its maximum, if other factors are non-limiting.
 WL_{opt} optimum water level for photosynthesis
 WL_{tol} water level tolerance, analogous to parameter T_{tol}
 a initial slope of the saturating GA response function

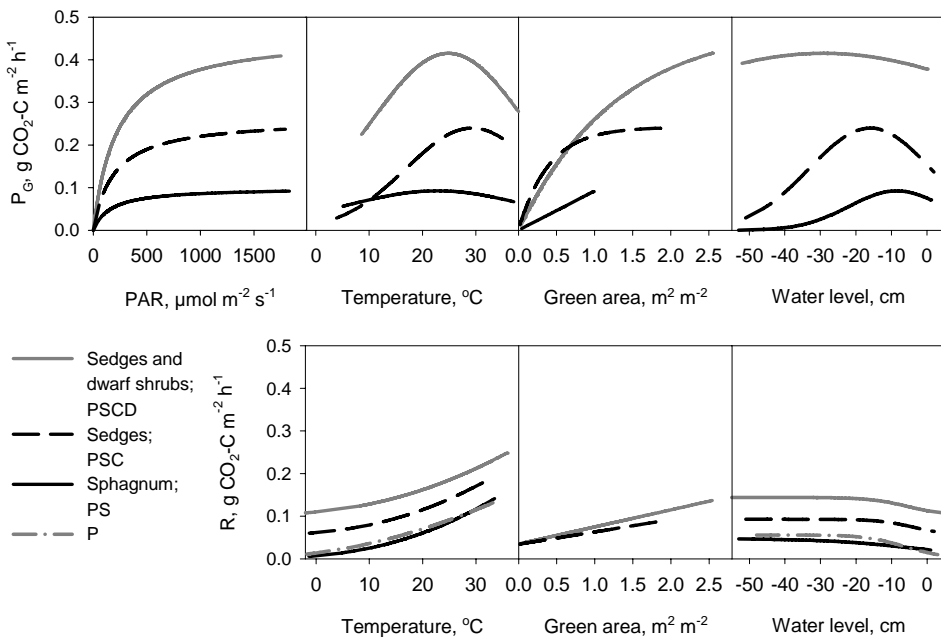


Figure 1. Modeled (Eqs. 2 and 3) response of gross photosynthesis (P_G) and respiration (R) to photosynthetically active radiation (PAR), air temperature, the amount of green area and water level. Water level is negative when it is below the moss/peat surface. The photosynthetic responses (upper panels) are shown for three plant groups: *Sphagnum* mosses, sedges, and mixed sedge and dwarf shrub vegetation. The respiratory responses (lower panels) are shown for different sample plot types: plots consisting of peat (P); peat and *Sphagnum* mosses (PS); peat, *Sphagnum* mosses and sedges (PSC); and peat, *Sphagnum* mosses, sedges and dwarf shrubs (PSCD). To illustrate the form of the responses, only one variable is allowed to change at the time while other are held constant. Data from Lakkasuo site, Paper I.

The photosynthetic response to PAR was described with a saturating function (term I) and the response to T and WL with unimodal (Gaussian) functions (terms II and III). In most cases, the response of photosynthesis to green area was saturating, described either with an exponential rise to maximum function (term IV, Paper I) or with a hyperbolic saturating function, similar to the response to PAR (Paper III). In case of *Sphagnum* mosses (Paper I) or plant communities where the green areas were quite low (Paper III), no self-shading could be observed and the response to GA was described as linear. In those cases, the P_{max} term has a slightly different interpretation: it denotes the potential maximum rate of gross photosynthesis when PAR, T and WL are non-limiting and GA (or the term s+GA, Paper III) equals 1. The form of the responses is illustrated in Fig. 1. In the plant community scale study (Paper III), the water level response (term III) could not be detected in two of the five communities.

$$R = R_{10} \exp \left[b_1 \left(\frac{1}{T_{ref} - T_0} - \frac{1}{T - T_0} \right) \right] \underbrace{\frac{1}{1 + \exp \left(- \frac{WL - b_2}{b_3} \right)}}_{II} + \underbrace{\frac{b_4 VGA}{III}}_{III} \quad (3)$$

The Eq. (3) has the following explanatory variables:

- T air temperature inside the measurement chamber (K)
- WL water level relative to the moss or peat surface (cm)
- VGA green area of vascular plants ($m^2 m^{-2}$)

And the following parameters:

- R_{10} respiration rate at 10 C, when WL is non-limiting and VGA is zero
- b_1 activation energy divided by the gas constant
- T_{ref} reference temperature, set at 283.15 K
- T_0 temperature minimum at which respiration reaches zero, set at 227.13 K (Lloyd and Taylor 1994)
- b_3 slope determining the speed and direction of change in R along the WL range
- b_2 WL at the centre of the fastest change along the WL range
- b_4 change in respiration per VGA unit

The response of R to T was exponential and described with a function from Lloyd & Taylor (1994) (term I). Air temperature inside the measurement chamber and soil temperatures in different depths were tested for the model; air temperature explained the variation in respiration best. The response to water level was described with a sigmoidal function (term II), except in the model for *Carex lasiocarpa* lawns (Paper III) where the water level gradient was not long enough for that parameterization and the form of exponential decay was used instead. The response to vascular green area was described as linear (term III). Term III was included when vascular plants were present. The form of the responses is illustrated in Fig. 1.

CO₂ exchange was reconstructed for the snow free season using the models (Papers I and III). CO₂ exchange during the snow covered season was reconstructed by linearly interpolating the fluxes between the winter measurement days (chamber or snow pack diffusion method) (Paper III). The last measurement at the end of and the first

measurement at the beginning of the snow-free season were used as a starting and end values for the interpolation.

2.3.2 *CH₄ flux measured using chamber method*

CH₄ fluxes were linearly interpolated between the measurement days to create a continuous time series (Papers **II** and **III**). The effect of linear interpolation was evaluated using the EC data and found to cause only a minor uncertainty in the seasonal flux estimate. Seven alternative sets of daily fluxes of every 7th day were extracted from the EC data and linearly interpolated to derive a seasonal estimate. These seven alternative estimates had a standard error of the mean that equaled 3% of the actual EC derived seasonal estimate (Paper **III**).

2.3.3 *Scaling up from the plant community level to the ecosystem*

In Siikaneva, one of the aims was to study the effect of the spatial variation in the plant community level on the ecosystem C gas fluxes (Paper **III**). Plant communities in the site were defined with TWINSpan cluster analysis, using the data from the vegetation inventory (Hill 1979, Hill and Šmilauer 2005). The analysis divided the vegetation into seven plant communities which we combined into five so that there were gas exchange sample plots in every community. The chamber measurements in different plant communities provided an estimate of the community specific C gas exchange. The proportion of each plant community within the ecosystem was determined based on the vegetation inventory. Community scale C gas exchange estimates were scaled to the ecosystem level by taking an area weighted mean. In case of CO₂, the differences between the vascular green leaf areas in the gas exchange sample plots and in the entire site were taken into account by reconstructing the CO₂ exchange using the corrected vascular green leaf areas. To test the effect of the fetch size on the results, the scaling was carried out for three observation radii (90 m, 120 m and 200 m radius from the EC mast). The upscaled estimates were compared with the EC derived estimates for the growing season 2005.

2.3.4 *Processing of the eddy covariance flux data*

The EC data acquisition was carried out with an in-house Python program BARFLUX (Finnish Meteorological Institute). The fluxes were calculated as half-hourly covariances (Eq. 1) using block averaging. Systematic flux losses were corrected for with empirical functions (Aubinet et al. 2000, Laurila et al. 2005). After data screening, 55% (CO₂) and 66% (CH₄) data coverage remained for the calculation of the annual balances. Gaps in the time series were filled using regression models (CO₂) and linear interpolation (CH₄).

3 RESULTS AND DISCUSSION

3.1 Carbon fluxes at the plant community component scale

3.1.1 CO_2

Photosynthesis at the plant community component scale was controlled by PAR, green area, temperature, and water level (Paper I). Based on the modeled responses, the optimum water level (parameter WL_{opt} in Eq. 2) for the photosynthesis of *Sphagnum* mosses, sedges, and mixed sedge and shrub vegetation of the lawn level plant community was 8.5, 16, and 29 cm below the moss surface, respectively (Fig. 1, Paper I). The photosynthesis of *Sphagnum* mosses had the narrowest and sedges had intermediate tolerance to water level variation (parameter WL_{tol} in Eq. 2). The very wide tolerance range of the mixed sedge and shrub vegetation indicated that such vegetation photosynthesizes equally efficiently in practically any hydrological conditions observed in the study. This wide tolerance range reflects the genuine lack of sensitivity of the shrubs to water level variation (Bubier et al. 2003a) but also may indicate a wider tolerance of mixed vegetation than a vegetation population consisting of only one functional group. Because of the high level of noise in the data, it was not possible to estimate the photosynthesis rate and parameterize the photosynthesis model separately for shrubs by subtracting the modeled sedge and *Sphagnum* photosynthesis from the measured values in the plots with shrubs present.

Differences in the water level responses were reflected in the contribution of the groups to the whole community photosynthesis (Paper I). In hydrologically normal conditions, *Sphagnum* mosses were an important component of the plant community, contributing approximately 28% of the community photosynthesis. Their role was emphasized in the beginning and at the end of the growing season when the green area of the vascular plants was low (Fig. 1). However, because of the sensitivity of *Sphagna* to drought, their contribution was reduced to approximately 10% during water level drawdown conditions (Paper I). The results agree with earlier observations on the sensitivity of *Sphagnum* photosynthesis to moisture conditions (Silvola and Aaltonen 1984, Rydin and McDonald 1985a, Schipperges and Rydin 1998).

Both the annual (Paper I) and seasonal (Fig. 2) variation in the photosynthesis of sedges was mostly related to the variation in their green area, but water level conditions also had an impact. Sedge photosynthesis was slightly suppressed in water level drawdown conditions. Furthermore, as shrub photosynthesis increased in drier conditions, the relative contribution of sedges to the community photosynthesis changed from 60% in hydrologically normal conditions to 30% in water level drawdown conditions (Paper I).

Respiration at the plant community component scale was controlled by temperature, water level and vascular green area (Paper I). Because of the high level of noise in the respiration measurements, the models could not be parameterized for the different components, using the subtraction method, but they were parameterized by the plot types. All plot types showed similar temperature and water level responses.

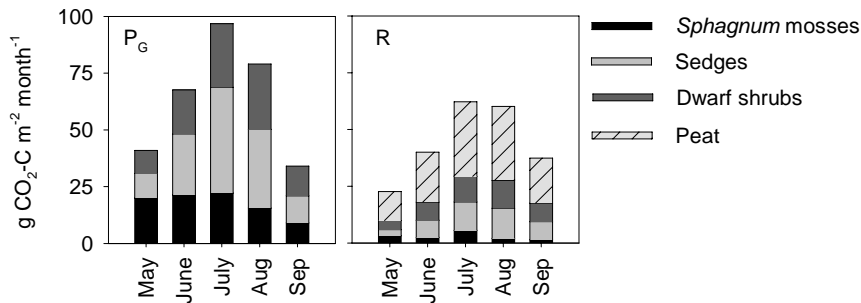


Figure 2. Monthly gross photosynthesis (P_G) and respiration (R) in Lakkasuo site, calculated as four year means. The fills in the bars show the contribution of each plant community component (*Sphagnum* mosses, sedges, dwarf shrubs and the underlying peat) to the total fluxes.

Both in Lakkasuo (Paper I) and Siikaneva (Paper IV), peat respiration accounted for approximately 50% of the total respiration in hydrologically normal conditions. Similar ratios between autotrophic and heterotrophic respiration have been obtained in earlier studies (Moore et al. 2002, Crow and Wieder 2005). Peat respiration and vascular plant respiration showed a clear seasonal pattern and their ratio was rather consistent throughout the growing season (Fig. 2). Relative to the amount of leaf area, shrubs had a higher respiration rate than sedges (Paper I), woody tissue constituting a significant proportion of the shrub respiration (Levy and Jarvis 1998, Saveyn et al. 2008). Peat respiration increased in water level drawdown conditions, while plant-derived respiration showed little variation between conditions (Papers I and IV).

3.1.2 Methane

Sedges were the most important plant group in regulating CH_4 fluxes (Paper II). In hydrologically normal conditions, the presence of sedges had a major impact increasing the CH_4 fluxes. CH_4 emissions were more than twice as high in the plots where sedges were present than in the plots where they had been removed. Similarly, in other sedge dominated sites, plant-mediated flux has been observed to account for 75 to 97% of the total flux (Whiting and Chanton 1992, Schimel 1995, Kelker and Chanton 1997). However, only the presence, rather than the number of sedges was significant within the range of sedge green leaf areas observed in this study (Fig. 3).

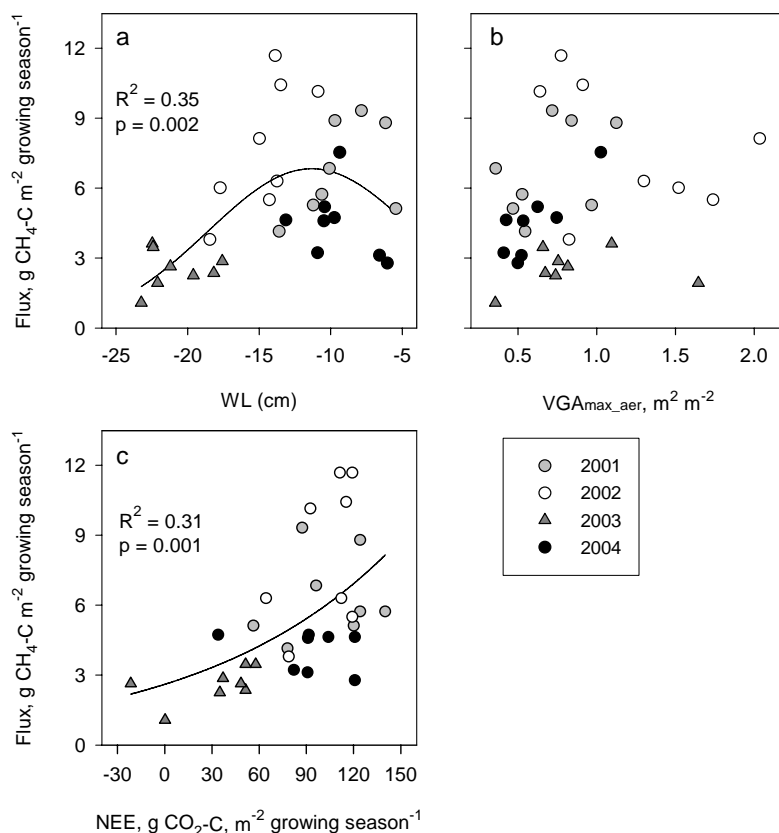


Figure 3. The relationship between the plot-wise seasonal (June-September) CH_4 emission and **a)** seasonal mean water level (WL), **b)** peak season vascular green area of aerenchymatous plants ($\text{VGA}_{\text{max_aer}}$) and **c)** seasonal net ecosystem CO_2 exchange (NEE). Water level is negative when it is below the moss surface. NEE is positive when the system is a sink of atmospheric CO_2 . In **a)** a Gaussian and in **b)** an exponential curve was fitted, using data points from all years. R^2 and p are the coefficient of determination and the significance level of the regression, respectively. Data from Lakkasuo control subsite, plots with unmanipulated vegetation.

The vegetation removal treatments suggested that the presence of shrubs may have a slightly attenuating effect on CH_4 fluxes (Paper II). The possible reasons behind these indicative results are the decreased substrate supply to the anaerobic decomposition chain due to the increased biomass allocation of sedges from the roots to the shoots in the presence of shrubs (Aerts et al. 1991), and the chemical suppression of methane fluxes by certain lignin compounds (Yavitt et al. 2000) that might be linked to the presence of shrubs (Paper II). However, more research on this topic needed.

During water level drawdown, the presence or absence of the vegetation components had no effect on the fluxes (Paper **II**), supporting the findings by Waddington and others (1996) and Strack and others (2006b). The fluxes were equally low in all vegetation treatment types. Only a small fraction of the sedge roots reach the CH₄ production zone in water level drawdown conditions. As a result, most of the recently fixed carbon, which is a more easily decomposable substrate for the anaerobic decomposition chain than the old, recalcitrant peat, is not available for CH₄ production. Water level acts as a switch. Only after passing through the water level control can the vegetation composition have an impact on the fluxes.

3.2 Carbon fluxes at the plant community scale

3.2.1 CO₂

The plant communities in Siikaneva differed in their CO₂ exchange (Paper **III**). Photosynthesis varied more between the communities than respiration, thus controlling the variation in NEE. *Carex lasiocarpa* lawns and hummocks had the highest, *Eriophorum vaginatum* lawns and *Carex rostrata* lawns had intermediate and hollows had the lowest NEE. This pattern is similar to the pattern in peak season VGAs. Besides spatial variation, temporal variation in the fluxes was considerable. The four-year study in Lakkasuo showed marked annual variation in the fluxes. The growing season NEE averaged 120 g CO₂-C during the years 2001, 2002 and 2004, but only 34 g CO₂-C during the dry year 2003 (Paper **I**). The CO₂ exchange estimates in Lakkasuo in hydrologically normal conditions were similar to the estimates in the Siikaneva lawn level communities. The experimental water level drawdown in Lakkasuo (Paper **I**) and the drought period in Siikaneva (Paper **IV**) caused a decrease in photosynthesis and an increase in respiration, resulting in a reduced CO₂ sink function.

3.2.2 Methane

The CH₄ emissions from the Siikaneva plant communities ranged from 6 (hummocks) to 19 (*Carex lasiocarpa* lawns) g CH₄-C growing season⁻¹ (June-September) (Paper **III**), and they were slightly higher in 2004 than in 2005. In Lakkasuo, the seasonal CH₄ emission in the control site was on average 6.5 g CH₄-C growing season⁻¹ during the hydrologically normal years (2001, 2002 and 2004) and only 2.5 g CH₄-C growing season⁻¹ during the dry year 2003. Although the CO₂ fluxes in Lakkasuo site were similar to those in Siikaneva lawns, they showed lower CH₄ fluxes, despite their similar VGAs and WLs. This demonstrates how large the site-to-site variation can be, even between peatlands located in the same region. Spatial variation in CH₄ fluxes was related to water level and NEE (Fig. 3). The water level response was unimodal, similar to Strack et al. (2004). In peatlands, the plant communities in the wettest microforms are often associated with the sparsest vascular plant cover and lowest net CO₂ uptake rate (Paper **III**, Waddington and Roulet 2000, Laine et al. 2007b). Thus, less substrate for CH₄ production is available in those communities.

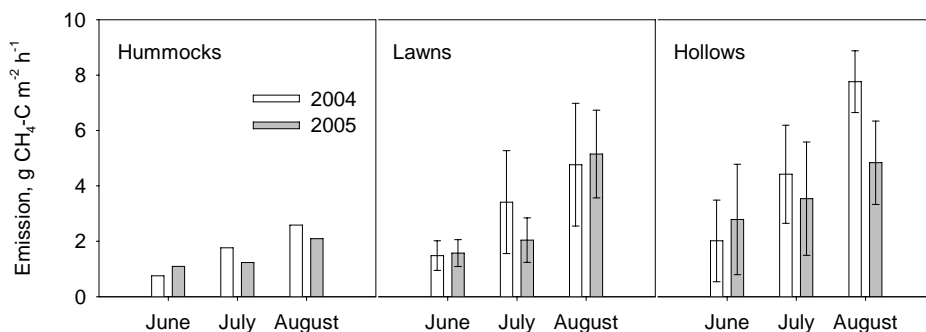


Figure 4. Mean measured methane emission in hummocks, lawns and hollows in Siikaneva site in mid-June, Mid-July and Mid-August. Water level during the drought period in July 2005 was 20 cm lower than in July 2004.

Both the experimental water level drawdown and natural droughts had an attenuating effect on the fluxes. The experimental water level drawdown in Lakkasuo significantly decreased the CH₄ fluxes, almost to zero (Paper II). In Siikaneva, the fluxes in all microforms were lower during the dry July 2005 than during the wet July 2004, while the differences were microform specific during the other months (Fig. 4).

3.3 Carbon fluxes at the ecosystem scale

3.3.1 CO₂

Siikaneva was a net sink of atmospheric CO₂ during the years 2004 and 2005 taking up, on average, 56 g CO₂-C a⁻¹ (Paper IV). The estimate agrees with the observations from other sedge dominated fens (Table 1). The seasonal cycle in the CO₂ exchange was considerable. The fen acted as a CO₂ sink during the five summer months (May-September) and as a small source of CO₂ to the atmosphere during the rest of the year. CO₂ fluxes had a clear diurnal pattern during the snow free season that followed the radiation conditions (Paper IV). Winter fluxes showed little variation (Paper IV). Their contribution to the annual NEE was, however, significant, approximately 25%.

By combining the photosynthetic and respiratory water level responses, it was concluded that the optimum water level for NEE in Siikaneva at the ecosystem scale was approximately -10 cm (Paper IV). This value matches the growing season average water level (-10 cm in 2004 and -13 in 2005), calculated from the water level measurements in the gas exchange sample plots. This optimum water level in Siikaneva is, interestingly, similar to that in an Irish blanket bog (Laine et al. 2007a) and in a Swedish poor fen (Yurova et al. 2007).

During July 2005 there was a short drought period, when the water level dropped to 25 cm below the surface. This deviation from the optimum water level was reflected as lower NEE (i.e. weaker net CO₂ sink) (Paper IV). The increase in respiration, mainly peat

respiration, was the main factor in the lowered NEE, but also photosynthesis was slightly suppressed. Despite the short drought in 2005, the annual balances were quite similar during both study years (Paper IV).

3.3.2 Methane

In Siikaneva, the annual CH₄ emission, measured by EC, was 9.4 g CH₄-C m⁻² (Paper V). The fluxes during the snow free period accounted for 91% of the annual flux. The annual and growing season CH₄ emissions from Siikaneva were typical for a northern fen (Table 1). CH₄ losses decreased the annual carbon balance estimate by 22% (Paper V).

The CH₄ fluxes showed a pronounced seasonal pattern (Papers III and V), but no diurnal variation (Paper V). Both the presence (Suyker et al. 1996, Juutinen et al. 2004) and the absence (Hyvönen et al. 1998, Kormann et al. 2001, Juutinen et al. 2004) of a diurnal cycle in wetlands has been previously observed. The diurnal cycle is strong in sites where the plant-mediated transport is based on pressured induced convection (Chanton et al. 1992, Kim et al. 1998). The CH₄ transport by *Carex* and *Eriophorum*, the dominant sedge genera in this study, is based on diffusion (Schimel 1995, Kutzbach et al. 2004). Besides transport through stomata that shows diurnal variation, cuticular conductance, not showing diurnal variation, is also an important mechanism of CH₄ release (Morrissey et al. 1993, Greenup et al. 2000). This would explain the absence of a diurnal pattern in Siikaneva.

Temporal variation in CH₄ fluxes at the daily scale was mainly controlled by peat temperature (Paper V). The temperature relationship was rather peculiar: no relationship could be seen at temperatures > 12 °C (Paper V). These temperatures occurred from mid-July through August. A period of high fluxes that coincided with a rapid deepening of the water level was observed in mid-July. The change in water level probably caused a degassing of the methane pool in the soil. Simultaneously, the methane concentration in the soil, sampled using a semi-permeable silicon tubes, showed a marked decrease (Riutta et al. 2007).

Daily variation in CH₄ fluxes during the year 2005 was not controlled by the water level (Paper V) and the between-community variation in the fluxes was not directly linked to the water level of the community, either (Paper III). Nevertheless, the CH₄ emissions were lower during the drought period in July 2005 than during the wet July 2004 (Fig. 4). Treat and others (2007) found that the relationships between CH₄ fluxes and controlling factors are clearest in monthly, rather than daily, time scales, because time-averaging reduces noise in the data. Such analysis, however, requires multi-year data sets that were not yet available in Siikaneva site. Within a lawn community in Lakkasuo, the seasonal average water level and CH₄ flux had a relationship (Fig. 3). However, the water level explained the year-to-year variation in the fluxes rather than the spatial variation within a given year.

3.4 Scaling up from the plant community level to the ecosystem

As is typical of fen ecosystems, both the microtopographical variation and the variation in plant community composition in Siikaneva were modest (Paper III). The fen was dominated by lawn vegetation. Some hummocks and hollows were also present. Despite the modest variation in microtopography and vegetation, however, the study on the spatial variation on the C gas fluxes showed that plant communities differed markedly in their C gas exchange (Paper III). Consequently, to provide C gas exchange estimates for the entire

fen ecosystem, it was important to take the spatial variation in fluxes and in vegetation composition into consideration. The results indicate that, were the proportion of the plant communities to change in a changing climate, it would have important consequences for the carbon balance of the mire (Paper **III**).

The chamber based estimates for the growing season NEE and CH₄ emission were in good agreement with the EC derived estimates (Paper **III**). However, the annual estimates differed more between the two methods. Outside the growing season, the sporadic measurements by the chamber and snowpack diffusion techniques were not representative for the entire autumn and winter period. The frequency of the winter measurements by the chamber and snow pack diffusion techniques was insufficient for the proper estimation of the winter time emissions. Rather, the measurements serve as an estimate of the magnitude of the fluxes during winter. Therefore, the annual fluxes estimated by the EC method (Paper **IV**, Paper **V**) are more reliable than those estimated by the chamber method (Paper **III**).

3.5 Synthesis of the water level drawdown impact

The experimental water level drawdown (Lakkasuo) and natural droughts (Lakkasuo during the growing season 2003 and Siikaneva during July 2005) caused changes in the carbon gas exchange. Of the plant community components, *Sphagnum* moss photosynthesis clearly decreased, sedge photosynthesis slightly decreased and dwarf shrub photosynthesis increased during water level drawdown (Fig. 5, Paper **I**). Because the responses of the components were compensatory, changes in photosynthesis and NEE were smaller at the plant community scale than at the plant community component scale (Fig. 5, Paper **I**). The results indicate that functional diversity buffers the ecosystem against environmental variability. If the drier conditions persist, the species abundance may gradually shift from sedge and *Sphagnum* dominance towards dwarf shrub dominance. Studies on the effects of the long-term water level draw-down (forestry drainage) on vegetation indicate that at the first stage of the drawdown induced succession, sedges decline and shrubs become more abundant (Laiho et al. 2003).

The water level drawdown resulted in a smaller CO₂ sink function, due to the decrease in photosynthesis and increase in respiration (Fig. 5, Papers **I** and **IV**). Because of the complementary responses of the plant groups, the reduced CO₂ sink function was mainly caused by the increase in peat respiration (Papers **I** and **IV**).

CH₄ fluxes decreased during the natural droughts and as a result of the water level drawdown treatment (Fig. 4, Fig. 5, Paper **II**). The moderate and short drought in Siikaneva during July 2005 had less impact on the fluxes than the long and severe drought in Lakkasuo from mid-June to September 2003. (Fig. 5). In Lakkasuo, the three-year experimental water level drawdown had a stronger impact than the natural drought. While CH₄ oxidation responds to environmental changes within a few days (Blodau and Moore 2003), CH₄ production recovers more slowly after a drought period, with an equilibration time of months (Freeman et al. 2002, Blodau and Moore 2003).

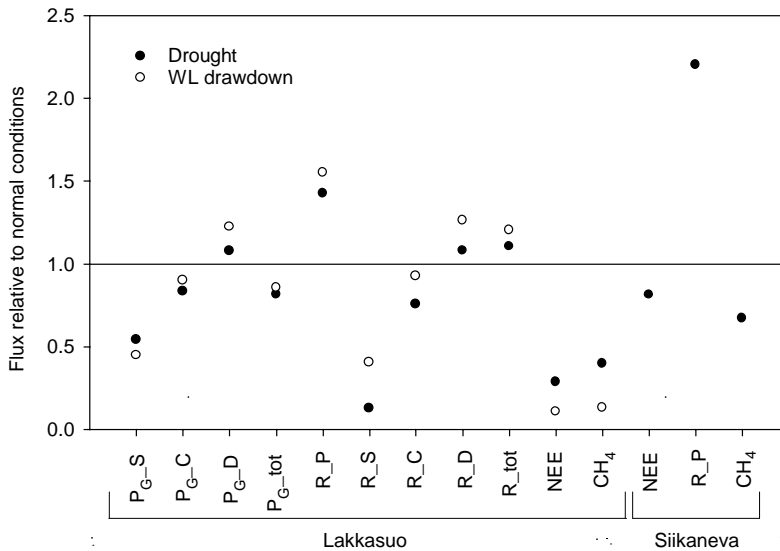


Figure 5. Impact of natural drought and experimental water level (WL) drawdown on photosynthesis (P_G), respiration (R), net ecosystem CO₂ exchange (NEE), and CH₄ emissions. The impact of the natural drought on the CO₂ and CH₄ fluxes in Lakkasuo is expressed as the ratio between the fluxes during the dry season 2003 and the average of the hydrologically normal seasons 2001, 2002 and 2004 in the control subsite. The impact of the experimental WL drawdown on CO₂ fluxes in Lakkasuo is expressed as the ratio between the average simulated fluxes in prevailing WL conditions and in the 14 cm WL drawdown conditions. The impact of the experimental WL drawdown in CH₄ fluxes in Lakkasuo is expressed as the ration between the fluxes during the WL drawdown treatment years (2002-2004) in the control and water level drawdown subsites. The impact of the natural drought in Siikaneva is determined as the ratio between the fluxes during the dry July 2005 and the hydrologically normal July 2004. In Lakkasuo, the impact is shown for the different plant community components (S-Sphagnum mosses, C-sedges, D-dwarf shrubs and P-peat) as well as for the whole community (tot). In Siikaneva, the NEE is measured by the eddy covariance technique and the peat respiration and CH₄ fluxes by chambers.

Deeper water levels do not necessarily stimulate CH₄ oxidation but the decrease in fluxes occurs mainly through changes in CH₄ production (Freeman et al. 2002) or transport. Furthermore, sulfate availability in the pore water increases during the low water level regime (Freeman et al. 1993, Dowrick et al. 2006), benefiting sulfate-reducing microorganisms that may out-compete methanogens for substrate (Yavitt and Lang 1990). Strack and others (2004, 2006b, 2007) showed that the water level response depends on the initial water level and on the plant community composition. For example, inundated hollows with sparse vascular cover may be colonized by sedges in drier conditions, causing no changes or an increase in CH₄ emissions (Strack et al. 2006b). This is in agreement with the Siikaneva results that the CH₄ fluxes are similar from lawns and hollows, the former having denser sedge cover and NEE and the latter having a higher water level and, therefore, larger CH₄ production zone (Paper III).

The results of this study examine the short term responses of a fen ecosystem and its components to water level drawdown. The results are applicable to climatic situations where droughts become more frequent but wet periods are still frequent enough as to maintain the fen vegetation composition relatively unchanged, consisting of *Sphagnum* mosses, sedges and dwarf shrubs. In case of a permanent water level drawdown, the changes in vegetation composition will mediate the changes in the carbon gas dynamics. The photosynthesis rate may recover when species adapted to drier conditions become more abundant and the decomposition rate may slow down due to the more recalcitrant litter (Laiho 2006). Therefore, the results of this study do not reflect the peatland carbon cycle in permanently drier climate.

3.6 Uncertainty in the estimates

The uncertainty in the flux measurements is considerable. In Siikaneva, the uncertainty in the EC-derived annual NEE and CH₄ emissions was estimated to be 25 and 3% for CO₂ and CH₄, respectively (Papers IV and V). The uncertainty analysis of the CO₂ fluxes considered more sources of error than that of the CH₄ fluxes and it is therefore a more realistic estimate of the magnitude of the uncertainty. The uncertainty was related to the instrumental errors, chosen filtering criteria, and gap-filling technique. Baldocchi (2003) estimated the error in EC derived annual NEE to be less than 50 g C m⁻² when measurements come from nearly ideal sites. This is a high figure relative to the typical annual NEE in fens (Table 1).

The uncertainty in the chamber derived estimates was even larger because of the limited spatial and temporal coverage and uncertainty in the temporal interpolation and spatial upscaling that were needed to estimate the ecosystem scale fluxes (Paper III). Both the reconstruction of the continuous time series and the upscaling caused considerable uncertainty in the estimates (Table 3 in Paper III). In addition, spatial variation, an inherent characteristic of a peatland ecosystem, was a source of further uncertainty. The total uncertainty in the seasonal NEE and CH₄ emissions was estimated to be 40% and 15%, respectively (Paper III). The uncertainty analysis did not take into account all sources of error, for example instrumental uncertainties. These values should, therefore, be only considered as indicative.

In Lakkasuo, the main aim was to reliably determine the impact of the water level drawdown, rather than to derive as accurate CO₂ exchange estimates as possible. As the results on the response of CO₂ exchange to water level drawdown were based largely on modeling, the uncertainty in the results that was related to the uncertainty in the model parameters was assessed (Paper I). The exercise showed that the impact of the water level drawdown and the role of the plant community components were evident enough as not to be obscured by the small changes in the model parameter values. In the Lakkasuo study (Papers I and II), the largest source of uncertainty is related to the vegetation removal treatments: the manipulations can never fully represent the vegetation components of an intact ecosystem. However, the results are consistent with the previous observations on the role of peat respiration (Crow and Wieder 2005), the sensitivity of *Sphagnum* mosses to variation in moisture conditions (e.g. Rydin and McDonald 1985b), the response of sedges and shrubs to drought (Bubier et al. 2003b), and the role of vegetation in CH₄ fluxes under different water level regimes (Waddington et al. 1996). Therefore, the results can be considered at least qualitatively reliable.

4 CONCLUSIONS

Fen ecosystem CO₂ and CH₄ dynamics during changing hydrological conditions was studied at different spatial scales, namely plant community component (*Sphagnum* mosses, sedges, dwarf shrubs and the underlying peat), plant community (hummocks, lawns and hollows) and ecosystem scale. The study shows that vegetation composition and water level impact are closely coupled. The response of the system to variations in hydrological conditions depends on the plant community composition. The photosynthesis of *Sphagnum* mosses and sedges decreases during water level drawdown conditions whereas the photosynthesis of dwarf shrubs increases. Consequently, functional diversity buffers the ecosystem against environmental variability. Because spatial variation between in C gas dynamics between the plant communities is considerable, their proportions determine the ecosystem scale fluxes. Thus, a shift in the community proportions during changing climatic conditions modifies the ecosystem fluxes.

The water level drawdown has a dual impact on the peatland ecosystem, and as a result, on the climate system. In water level drawdown conditions, the fen ecosystem becomes a smaller CO₂ sink and smaller CH₄ source to the atmosphere. However, if the drier conditions persist, changes in vegetation composition will mediate the changes in the carbon gas dynamics. Therefore, the results of this study are applicable to the short term impacts of the water level drawdown and to climatic situations where droughts become more frequent but there are still enough wet periods to maintain the fen vegetation composition relatively unchanged, consisting of *Sphagnum* mosses, sedges and dwarf shrubs.

REFERENCES

- Aerts, R., Boot, R. G. A. & Vandraart, P. J. M. 1991. The relation between aboveground and belowground biomass allocation patterns and competitive ability. *Oecologia* 87: 551-559.
- Aguiar, M. R., Paruelo, J. M., Sala, O. E. & Lauenroth, W. K. 1996. Ecosystem responses to changes in plant functional type composition: An example from the Patagonian steppe. *Journal of Vegetation Science* 7: 381-390.
- Ahti, T., Hämet-Ahti, L. & Jalas, J. 1968. Vegetation zones and their sections in northwestern Europe. *Annales Botanici Fennici* 5: 169-211.
- Alm, J., Saarnio, S., Nykänen, H., Silvola, J. & Martikainen, P. J. 1999a. Winter CO₂, CH₄ and N₂O fluxes on some natural and drained boreal peatlands. *Biogeochemistry* 44: 163-186.
- , Schulman, L., Walden, J., Nykänen, H., Martikainen, P. J. & Silvola, J. 1999b. Carbon balance of a boreal bog during a year with an exceptionally dry summer. *Ecology* 80: 161-174.
- , Shurpali, N. J., Tuittila, E. S., Laurila, T., Maljanen, M., Saarnio, S. & Minkkinen, K. 2007. Methods for determining emission factors for the use of peat and peatlands - flux measurements and modelling. *Boreal Environment Research* 12: 85-100.

- , Talanov, A., Saarnio, S., Silvola, J., Ikkonen, E., Aaltonen, H. et al. 1997. Reconstruction of the carbon balance for microsites in a boreal oligotrophic pine fen, Finland. *Oecologia* 110: 423-431.
- Andreev, A. A. & Klimanov, V. A. 2000. Quantitative Holocene climatic reconstruction from Arctic Russia. *Journal of Paleolimnology* 24: 81-91.
- Antonsson, K. & Seppä, H. 2007. Holocene temperatures in Bohuslän, southwest Sweden: a quantitative reconstruction from fossil pollen data. *Boreas* 36: 400 – 410.
- Aubinet, M., Grelle, A., Ibrom, A., Rannik, U., Moncrieff, J., Foken, T. et al. 2000. Estimates of the annual net carbon and water exchange of forests: The EUROFLUX methodology. *Advances in Ecological Research* 30: 113-175.
- Aurela, M., Laurila, T. & Tuovinen, J. P. 2004. The timing of snow melt controls the annual CO₂ balance in a subarctic fen. *Geophysical Research Letters* 31: L16119. doi: 10.1029/2004GL020315.
- Baldocchi, D. D. 2003. Assessing the eddy covariance technique for evaluating carbon dioxide exchange rates of ecosystems: past, present and future. *Global Change Biology* 9: 479-492.
- Bedford, B. L., Rappaport, N. R. & Bernard, J. M. 1988. A life history of *Carex lasiocarpa* Ehrh ramets. *Aquatic Botany* 30: 63-80.
- Bellisario, L. M., Bubier, J. L., Moore, T. R. & Chanton, J. P. 1999. Controls on CH₄ emissions from a northern peatland. *Global Biogeochemical Cycles* 13: 81-91.
- Bergman, I., Svensson, B. H. & Nilsson, M. 1998. Regulation of methane production in a Swedish acid mire by pH, temperature and substrate. *Soil Biology & Biochemistry* 30: 729-741.
- Bernard, J. M. 1976. Life history and population dynamics of shoots of *Carex rostrata*. *Journal of Ecology* 64: 1045-1048.
- , Solander, D. & Kvet, J. 1988. Production and nutrient dynamics in *Carex* wetlands. *Aquatic Botany* 30: 125-147.
- Blodau, C. & Moore, T. R. 2003. Experimental response of peatland carbon dynamics to a water table fluctuation. *Aquatic Sciences* 65: 47-62.
- Botch, M. S. & Masing, V. V. 1983. Mire ecosystem in the U.S.S.R. In: Gore, A.J.P. (ed.). *Ecosystems of the world 4B mires: Swamp, bog, fen and moor. Regional studies.* Elsevier, Amsterdam. p. 95-152.
- Bremer, D. J., Ham, J. M. Owensby, C. E., Knapp, A. K. 1998. Responses of soil respiration to clipping and grazing in a tallgrass prairie. *Journal of Environmental Quality* 27: 1539-1548.
- Brix, H., Sorrell, B. K. & Orr, P. T. 1992. Internal pressurization and convective gas-glow in some emergent fresh-water macrophytes. *Limnology and Oceanography* 37: 1420-1433.
- Bubier, J. L., Bhatia, G., Moore, T. R., Roulet, N. T. & Lafleur, P. M. 2003a. Spatial and temporal variability in growing-season net ecosystem carbon dioxide exchange at a large peatland in Ontario, Canada. *Ecosystems* 6: 353-367.
- , Crill, P., Mosedale, A., Frolking, S. & Linder, E. 2003b. Peatland responses to varying interannual moisture conditions as measured by automatic CO₂ chambers. *Global Biogeochemical Cycles* 17: 1066. doi:10.1029/2002GB001946
- Chanton, J. P., Martens, C. S., Kelley, C. A., Crill, P. M. & Shovers, W. J. 1992. Methane transport mechanisms and isotopic fractionation in emergent macrophytes of an Alaskan tundra lake. *Journal of Geophysical Research – Atmospheres* 97(D15): 16681-16688.
- Chapin, F. S., BretHarte, M. S., Hobbie, S. E. & Zhong, H. L. 1996. Plant functional types as predictors of transient responses of arctic vegetation to global change. *Journal of Vegetation Science* 7: 347-358.

- , Woodwell, G. M., Randerson, J. T., Rastetter, E. B., Lovett, G. M., Baldocchi, D. D. et al. 2006. Reconciling carbon-cycle concepts, terminology, and methods. *Ecosystems* 9: 1041-1050.
- Charman, D. 2002. Peatlands and environmental change. John Wiley & Sons, cop, Chichester. 301 p.
- Chasar, L. S., Chanton, J. P., Glaser, P. H., Siegel, D. I. & Rivers, J. S. 2000. Radiocarbon and stable carbon isotopic evidence for transport and transformation of dissolved organic carbon, dissolved inorganic carbon, and CH₄ in a northern Minnesota peatland. *Global Biogeochemical Cycles* 14: 1095-1108.
- Christensen, J. H., Hewitson, B., Busuioic, A., Chen, A., Gao, X., Held, I. et al. 2007. Regional climate projections. In: Solomon, S., Qin, D., Manning, M., Chen, Z., Marquis, M., Averyt, K.B. et al (eds.). *Climate change 2007: The physical science basis. Contribution of Working Group I to the fourth assessment report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge, UK. p. 847-940.
- Christensen, T. R., Ekberg, A., Strom, L., Mastepanov, M., Panikov, N., Mats, O. et al. 2003. Factors controlling large scale variations in methane emissions from wetlands. *Geophysical Research Letters* 30: 1414. doi:10.1029/2002GL016848.
- , Friborg, T., Sommerkorn, M., Kaplan, J., Illeris, L., Soegaard, H. et al. 2000. Trace gas exchange in a high-arctic valley 1. Variations in CO₂ and CH₄ flux between tundra vegetation types. *Global Biogeochemical Cycles* 14: 701-713.
- Clymo, R. S. 1983. Peat. In: Gore, A.J.P. (ed.). *Ecosystems of the of the world 4A mires: Swamp, bog, fen and moor. General studies*. Elsevier, Amsterdam. p. 159-224.
- 1984. The limits to peat bog growth. *Philosophical Transactions of the Royal Society of London Series B-Biological Sciences* 303: 605-654.
- & Hayward, P. M. 1982. The ecology of Sphagnum. In: Smith, A.J.E. (ed.). *Bryophyte ecology*. Chapman and Hall, London. p. 229-289.
- & Reddaway, E. J. F. 1971. Productivity of Sphagnum (bogmoss) and peat accumulation. *Hydrobiologia* 12: 181-192.
- , Turunen, J. & Tolonen, K. 1998. Carbon accumulation in peatland. *Oikos* 81: 368-388.
- Crill, P. M., Bartlett, K. B., Harriss, R. C., Gorham, E. & Verry, E. S. 1988. Methane flux from Minnesota peatlands. *Global Biogeochemical Cycles* 2: 371-384.
- Crow, S. E. & Wieder, R. K. 2005. Sources of CO₂ emission from a northern peatland: Root respiration, exudation, and decomposition. *Ecology* 86: 1825-1834.
- Dacey, J. W. H. & Klug, M. J. 1979. Methane efflux from lake sediments through water lilies. *Science* 203: 1253-1255.
- Daulat, W. E. & Clymo, R. S. 1998. Effects of temperature and water table on the efflux of methane from peatland surface cores. *Atmospheric Environment* 32: 3207-3218.
- Denman, K. L., Brasseur, G., Chidthaisong, A., Ciais, P., Cox, P. M., Dickinson, R. E. et al. 2007. Couplings between changes in the climate system and biogeochemistry. In: Solomon, S., Quin, D., Manning, M., Chen, Z., Marquis, M., Averyt, K.B. et al (eds.). *Climate change 2007: The physical science basis. Contribution of Working Group I to the fourth assessment report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.
- Dowrick, D. J., Freeman, C., Lock, M. A. & Reynolds, B. 2006. Sulphate reduction and the suppression of peatland methane emissions following summer drought. *Geoderma* 132: 384-390.
- Drebs, A., Nordlund, A., Karlsson, P., Helminen, J. & Rissanen, P. 2002. Climatological statistics of Finland 1971-2000. *Climatic Statistics of Finland* 2002/11-99.

- Dueck, T. A., de Visser, R., Poorter, H., Persijn, S., Gorissen, A., de Visser, W., Schapendonk, A., Verhagen, J., Snel, J., Harren, F.J. M., Ngai, A.K.Y., Verstappen, F., Bouwmeester, H., Voeseek, L. A. C. J. % van der Werf, A. 2007. No evidence for substantial aerobic methane emission by terrestrial plants: a ¹³C-labelling approach. *New Phytologist* 175: 29-35.
- Dunfield, P., Knowles, R., Dumont, R. & Moore, T. R. 1993. Methane production and consumption in temperate and sub-arctic peat soils - Response to temperature and pH. *Soil Biology & Biochemistry* 25: 321-326.
- Fagerstedt, K. V. 1992. Development of aerenchyma in roots and rhizomes of *Carex rostrata* (Cyperaceae). *Nordic Journal of Botany* 12: 115-120.
- Fechner-Levy, E. J. & Hemond, H. F. 1996. Trapped methane volume and potential effects on methane ebullition in a northern peatland. *Limology and Oceanography* 41: 1375-1383
- Fischlin, A., Midgley, G. F., Price, J. T., Leemans, R., Gopal, C., Turley, C. et al. 2007. Ecosystems, their properties, goods and services. In: Parry, M.L., Canziani, O.F., Palutikof, J.P., van der Linden, P.J. & Hanson, C.E. (eds.). *Climate change 2007: Impacts, adaptation and vulnerability. Contribution of Working Group II to the fourth assessment report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge, UK. p. 211-272.
- Francez, A. J. & Vasander, H. 1995. Peat accumulation and peat decomposition after human disturbance in French and Finnish mires. *Acta Oecologica* 16: 599-608.
- Freeman, C., Lock, M. A. & Reynolds, B. 1993. Fluxes of CO₂, CH₄ and N₂O from a Welsh peatland following simulation of water-table draw-down - Potential feedback to climatic change. *Biogeochemistry* 19: 51-60.
- , Nevison, G. B., Kang, H., Hughes, S., Reynolds, B. & Hudson, J. A. 2002. Contrasted effects of simulated drought on the production and oxidation of methane in a mid-Wales wetland. *Soil Biology & Biochemistry* 34: 61-67.
- Frenzel, P. & Rudolph, J. 1998. Methane emission from a wetland plant: the role of CH₄ oxidation in *Eriophorum*. *Plant and Soil* 202: 27-32.
- Frolking, S. & Roulet, N. T. 2007. Holocene radiative forcing impact of northern peatland carbon accumulation and methane emissions. *Global Change Biology* 13: 1079-1088.
- , Bubier, J. L., Moore, T. R., Ball, T., Bellisario, L. M., Bhardwa, A., Carroll, J. P., Crill, P. M., Lafleur, P. M., McCaughey, J. H., Roulet, N. T., Suyker, A. E., Verrna, S. B., Waddington, J. M. & Whiting, G. J. 1998. Relationship between ecosystem productivity and photosynthetically active radiation for northern peatlands. *Global Biogeochemical Cycles* 12: 115-126.
- Gaberscik, A. & Martincic, A. 1987. Seasonal dynamics of net photosynthesis and productivity of *Sphagnum papillosum*. *Lindbergia* 13: 105-110.
- Garnet, K. N., Magonigal, J. P., Litchfield, C. & Taylor, G. E. 2005. Physiological control of leaf methane emission from wetland plants. *Aquatic Botany* 81: 141-155.
- Gifford, R. M. 2003. Plant respiration in productivity models: conceptualisation, representation and issues for global terrestrial carbon-cycle research. *Functional Plant Biology* 30: 171-186.
- Gitay, H., Brown, S., Easterling, W., Jallow, B., Antle, J., Apps, M. et al. 2001. Ecosystems and their goods and services. In: McCarthy, J.J., Canziani, O.F., Leary, N.A., Dokken, D.J. & White, K.S. (eds.). *Climate change 2001: Impacts, adaptation and vulnerability. Contribution of Working Group II to the third assessment report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge, United Kingdom. p. 235-342.

- Glaser, P. H., Chanton, J. P., Morin, P., Rosenberry, D. O., Siegel, D. I., Ruud, O., Chasar, L. I. & Reeve, A. S. 2004. Surface deformations as indicators of deep ebullition fluxes in a large northern peatland. *Global Biogeochemical Cycles* 18: GB1003. doi:10.1029/2003GB002069.
- Gorham, E. 1991. Northern peatlands - Role in the carbon cycle and probable responses to climatic warming. *Ecological Applications* 1: 182-195.
- Greenup, A. L., Bradford, M. A., McNamara, N. P., Ineson, P. & Lee, J. A. 2000. The role of *Eriophorum vaginatum* in CH₄ flux from an ombrotrophic peatland. *Plant and Soil* 227: 265-272.
- Griffis, T. J. & Rouse, W. R. 2001. Modelling the interannual variability of net ecosystem CO₂ exchange at a subarctic sedge fen. *Global Change Biology* 7: 511-530.
- Hargreaves, K. J., Fowler, D., Pitcairn, C. E. R. & Aurela, M. 2001. Annual methane emission from Finnish mires estimated from eddy covariance campaign measurements. *Theoretical and Applied Climatology* 70: 203-213.
- Harley, P. C., Tenhunen, J. D., Murray, K. J. & Beyers, J. 1989. Irradiance and temperature effects on photosynthesis of tussock tundra *Sphagnum* mosses from the foothills of the Philip Smith Mountains, Alaska. *Oecologia* 79: 251-259.
- Heikkinen, J. E. P., Elsakov, V. & Martikainen, P. J. 2002. Carbon dioxide and methane dynamics and annual carbon balance in tundra wetland in NE Europe, Russia. *Global Biogeochemical Cycles* 16: 1115. doi:10.1029/2002GB001930
- Hill, M. O. 1979. TWINSPLAN – a FORTRAN program for arranging multivariate data in an ordered two-way table by classification of the individuals and attributes. *Ecology and Systematics*, Cornell University, Ithaca, New York.
- & Šmilauer, P. 2005. TWINSPLAN for Windows version 2.3. Centre for Ecology and Hydrology & University of South Bohemia, Huntingdon & Ceske Budejovice. 29 p.
- Hyvönen, T., Ojala, A., Kankaala, P. & Martikainen, P. J. 1998. Methane release from stands of water horsetail (*Equisetum fluviatile*) in a boreal lake. *Freshwater Biology* 40: 275-284.
- IPCC. 2007. Summary for policymakers. In: Solomon, S., Qin, D., Manning, M., Chen, Z., Marquis, M., Averyt, K.B. et al (eds.). *Climate change 2007: The physical science basis. Contribution of Working Group I to the fourth assessment report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge, United Kingdom. p. 1-18.
- Jeglum, J. K. 1974. Relative influence of moisture-aeration and nutrients on vegetation and black spruce growth in northern Ontario. *Canadian Journal of Forest Research* 4: 114-126.
- Joabsson, A., Christensen, T. R. & Wallen, B. 1999. Vascular plant controls on methane emissions from northern peatforming wetlands. *Trends in Ecology & Evolution* 14: 385-388.
- Joosten, H. & Clarke, D. 2002. *Wise use of mires and peatlands: Background and principles including a framework for decision-making*. International Mire Conservation Group and International Peat Society. Jyväskylä. 300 p.
- Juutinen, S., Alm, J., Larmola, T., Saarnio, S., Martikainen, P. J. & Silvola, J. 2004. Stand-specific diurnal dynamics of CH₄ fluxes in boreal lakes: Patterns and controls. *Journal of Geophysical Research – Atmospheres* 109(D19): D19313. doi:10.1029/2004JD004782
- Karlsson, P. S. 1989. In situ photosynthetic performance of four coexisting dwarf shrubs in relation to light in a subarctic woodland. *Functional Ecology* 3: 481-487.
- Kelker, D. & Chanton, J. 1997. The effect of clipping on methane emissions from *Carex*. *Biogeochemistry* 39: 37-44.

- Keller, J. K. & Bridgman, S. E. 2007. Pathways of anaerobic carbon cycling across an ombrotrophic-minerotrophic peatland gradient. *Limnology and Oceanography* 52: 96-107.
- Kepler F., Hamilton, J. T.G., Brass, M. & Röckmann, T. 2006. Methane emissions from terrestrial plants under aerobic conditions. *Nature* 439: 187-191.
- , Röckmann, T., Vigano, I., McRoberts, C. & Hamilton, J. T. G. 2008. Methoxyl groups of plant pectin as a precursor compound for atmospheric methane: evidence from deuterium labelling studies. *Geophysical Research Abstracts* 10, EGU2008-A-02064. EGU General Assembly 2008.
- Kim, J., Verma, S. B., Billesbach, D. P. & Clement, R. J. 1998. Diel variation in methane emission from a midlatitude prairie wetland: Significance of convective through flow in *Phragmites australis*. *Journal of Geophysical Research – Atmospheres* 103(D21): 28029-28039.
- King, G. M. 1994. Associations of methanotrophs with the roots and rhizomes of aquatic vegetation. *Applied and Environmental Microbiology* 60: 3220-3227.
- King, J. Y., Reeburgh, W. S. & Regli, S. K. 1998. Methane emission and transport by arctic sedges in Alaska: Results of a vegetation removal experiment. *Journal of Geophysical Research – Atmospheres* 103(D22): 29083-29092.
- Korhola, A., Tolonen, K., Turunen, J. & Jugner, H. 1995. Estimating long-term carbon accumulation rates in boreal peatlands by radiocarbon dating. *Radiocarbon* 37: 575-584
- Kormann, R., Müller, H. & Werle, P. 2001. Eddy flux measurements of methane over the fen "Murnauer Moos", 11°11'E, 47°39'N, using a fast tunable diode laser spectrometer. *Atmospheric Environment* 35: 2533-2544.
- Kutzbach, L., Wagner, D. & Pfeiffer, E. M. 2004. Effect of microrelief and vegetation on methane emission from wet polygonal tundra, Lena Delta, Northern Siberia. *Biogeochemistry* 69: 341-362.
- Kuzyakov, Y. & Cheng, W. 2001. Photosynthesis controls of rhizosphere respiration and organic matter decomposition. *Soil Biology and Biochemistry* 33: 1915-1925.
- Lafleur, P. M., Roulet, N. T., Bubier, J. L., Frolking, S. & Moore, T. R. 2003. Interannual variability in the peatland-atmosphere carbon dioxide exchange at an ombrotrophic bog. *Global Biogeochemical Cycles* 17: 1036. doi:10.1029/2002GB001983.
- Laiho, R. 2006. Decomposition in peatlands: Reconciling seemingly contrasting results on the impacts of lowered water levels. *Soil Biology & Biochemistry* 38: 2011-2024.
- , Vasander, H., Penttilä, T. & Laine, J. 2003. Dynamics of plant-mediated organic matter and nutrient cycling following water-level drawdown in boreal peatlands. *Global Biogeochemical Cycles* 17: 1053. doi:10.1029/2002GB002015.
- Laine, A., Byrne, K. A., Kiely, G. & Tuittila, E. S. 2007a. Patterns in vegetation and CO₂ dynamics along a water level gradient in a lowland blanket bog. *Ecosystems* 10: 890-905.
- , Sottocornola, M., Kiely, G., Byrne, K. A., Wilson, D. & Tuittila, E. S. 2006. Estimating net ecosystem exchange in a patterned ecosystem: Example from blanket bog. *Agricultural and Forest Meteorology* 138: 231-243.
- , Wilson, D., Kiely, G. & Byrne, K. A. 2007b. Methane flux dynamics in an Irish lowland blanket bog. *Plant and Soil* 299: 181-193.
- Laine, J. & Vanha-Majamaa, I. 1992. Vegetation ecology along a trophic gradient on drained pine mires in southern Finland. *Annales Botanici Fennici* 29: 213-233.
- , Silvola, J., Tolonen, K., Alm, J., Nykänen, H., Vasander, H., Sallantausta, T., Savolainen, I., Sinisalo, J. & Martikainen, P. J. 1996. Effect of water-level drawdown on global climatic warming: Northern peatlands. *Ambio* 25: 179-184.

- , Vasander, H. & Laiho, R. 1995. Long-term effects of water level drawdown on the vegetation of drained pine mires in southern Finland. *Journal of Applied Ecology* 32: 785-802.
- Lappalainen, E. 1996. Global peat resources. International Peat Society and Geological Survey of Finland, Jyskä, Finland. 359 p.
- Laurila, T., Tuovinen, J. P., Lohila, A., Hatakka, J., Aurela, M., Thum, T. et al. 2005. Measuring methane emissions from a landfill using a cost-effective micrometeorological method. *Geophysical Research Letters* 32: L19808. doi:10.1029/2005GL023462.
- Le Mer, J. & Roger, P. 2001. Production, oxidation, emission and consumption of methane by soils: A review. *European Journal of Soil Biology* 37: 25-50.
- Levy, P. E. & Jarvis, P. G. 1998. Stem CO₂ fluxes in two Sahelian shrub species (*Guiera senegalensis* and *Combretum micranthum*). *Functional Ecology* 12: 107-116.
- Litton, C. M., Raich, J. W. & Ryan, M. G. 2007. Carbon allocation in forest ecosystems. *Global Change Biology* 13: 2089-2109.
- Lloyd, J. & Taylor, J. A. 1994. On the temperature-dependence of soil respiration. *Functional Ecology* 8: 315-323.
- Lovelock, C. E. & Feller, I. C. 2003. Photosynthetic performance and resource utilization of two mangrove species coexisting in a hypersaline scrub forest. *Oecologia* 134: 455-462.
- Lovley, D. R. & Klug, R. J. 1986. Model for the distribution of sulphate reduction and methanogenesis in freshwater sediments. *Geochimica et Cosmochimica Acta* 50: 11-18.
- , Coates, J. D., Blunt-Harris, E. L., Philips, E. J. P. & Woodward, J.C. 1996. Humic substances as electron acceptors for microbial respiration. *Nature* 382: 445-448.
- MacDonald, J. A., Fowler, D., Hargreaves, K. J., Skiba, U., Leith, I. D. & Murray, M. B. 1998. Methane emission rates from a northern wetland; Response to temperature, water table and transport. *Atmospheric Environment* 32: 3219-3227.
- Macdonald, S. E. & Yin, F. Y. 1999. Factors influencing size inequality in peatland black spruce and tamarack: evidence from post-drainage release growth. *Journal of Ecology* 87: 404-412.
- Malmer, N., Albinsson, C., Svensson, B. M. & Wallen, B. 2003. Interferences between Sphagnum and vascular plants: effects on plant community structure and peat formation. *Oikos* 100: 469-482.
- Matthews, E. & Fung, I. 1987. Methane emission from natural wetlands: Global distribution, area, and environmental characteristics of sources. *Global Biogeochemical Cycles* 1: 61-86.
- Mikaloff Fletcher, S. E., Tans, P. P., Bruhwiler, L. M., Miller, J. B. & Heimann, M. 2004. CH₄ sources estimated from atmospheric observations of CH₄ and its C-13. *Global Biogeochemical Cycles* 18: GB4004. doi:10.1029/2004GB002223.
- Mitchell, F. B. J. 1989. The “greenhouse” effect and climatic change- *Reviews in Geophysics* 27; 115– 139.
- Moog, P. R. & Bruggemann, W. 1998. Flooding tolerance of *Carex* species. II. Root gas-exchange capacity. *Planta* 207: 199-206.
- Moore, P. D. 2002. The future of cool temperate bogs. *Environmental Conservation* 29: 3-20.
- Moore, T. R. & Dalva, M. 1993. The influence of temperature and water table position on carbon dioxide and methane emissions from laboratory columns of peatland soils. *Journal of Soil Science* 44: 651-664.
- & Knowles, R. 1990. Methane emissions from fen, bog and swamp peatlands in Quebec. *Biogeochemistry* 11: 45-61.

- & Roulet, N. T. 1993. Methane flux - water table relations in northern wetlands. *Geophysical Research Letters* 20: 587-590.
- , Bubier, J. L., Frolking, S. E., Lafleur, P. M. & Roulet, N. T. 2002. Plant biomass and production and CO₂ exchange in an ombrotrophic bog. *Journal of Ecology* 90: 25-36.
- , Roulet, N. T. & Knowles, R. 1990. Spatial and temporal variations of methane flux from subarctic/northern boreal fens. *Global Biogeochemical Cycles* 4, 29-46.
- Morrissey, L. A., Zobel, D. B. & Livingston, G. P. 1993. Significance of stomatal control on methane release from *Carex*-dominated wetlands. *Chemosphere* 26: 339-355.
- Oechel, W. C., Vourlitis, G. L., Hastings, S. J. & Bochkarev, S. A. 1995. Change in arctic CO₂ flux over two decades - Effects of climate change at Barrow, Alaska. *Ecological Applications* 5: 846-855.
- Økland, R. H., Okland, T. & Rydgren, K. 2001. A Scandinavian perspective on ecological gradients in north-west European mires: reply to Wheeler and Proctor. *Journal of Ecology* 89: 481-486.
- Parsons, A. N., Welker, J. M., Wookey, P. A., Press, M. C., Callaghan, T. V. & Lee, J. A. 1994. Growth-responses of four sub-arctic dwarf shrubs to simulated environmental change. *Journal of Ecology* 82: 307-318.
- Pearce, D. M. E. & Clymo, R. S. 2001. Methane oxidation in a peatland core. *Global Biogeochemical Cycles* 15: 709-720.
- Pelletier, L., Moore, T. R., Roulet, N. T., Garneau, M. & Beaulieu-Audy, V. 2007. Methane fluxes from three peatlands in the La Grande Riviere watershed, James Bay lowland, Canada. *Journal of Geophysical Research -Biogeosciences* 112(G1): G01018. doi:10.1029/2006JG000216.
- Popp, T. J., Chanton, J. P., Whiting, G. J. & Grant, N. 1999. Methane stable isotope distribution at a *Carex* dominated fen in North Central Alberta. *Global Biogeochemical Cycles* 13: 1063-1078.
- , Chanton, J. P., Whiting, G. J. & Grant, N. 2000. Evaluation of methane oxidation in the rhizosphere of a *Carex* dominated fen in north central Alberta, Canada. *Biogeochemistry* 51: 259-281.
- Pumpanen, J., Kolari, P., Ilvesniemi, H., Minkkinen, K., Vesala, T., Niinistö, S. et al. 2004. Comparison of different chamber techniques for measuring soil CO₂ efflux. *Agricultural and Forest Meteorology* 123: 159-176.
- Ramaswamy, V., Boucher, O., Haigh, J., Hauglustaine, D., Haywood, J., Myhre, G. et al. 2001. Radiative forcing of climate change. In: Houghton, J.T., Ding, Y., Griggs, D.J., Noguer, M., van der Linden, P.J., Dai, X. et al (eds.). *Climate change 2001: The scientific basis. Contribution of Working Group I to the third assessment report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge, United Kingdom. p. 349-416.
- Rinnan, R., Impio, M., Silvola, J., Holopainen, T. & Martikainen, P. J. 2003. Carbon dioxide and methane fluxes in boreal peatland microcosms with different vegetation cover - effects of ozone or ultraviolet-B exposure. *Oecologia* 137: 475-483.
- Riutta, T., Juottonen, H., Rinne, J., Juutinen, S., Pumpanen, J., Aurela, M., Laine, J., Putkinen, A., Yrjälä, K., Vesala, T., Hari, P., Laurila, T., Pihlatie, M., Haapanala, S., Tuittila, E. 2007. Methane dynamics in a boreal peatland: combining flux measurements, concentration profiles in the peat and microbial ecology at different spatial and temporal scales. *Eos Trans. AGU*, 88(52), Fall Meet. Suppl., Abstract B53A-0924.
- Roulet, N. T., Lafleur, P. M., Richard, P. J. H., Moore, T. R., Humphreys, E. R. & Bubier, J. 2007. Contemporary carbon balance and late Holocene carbon accumulation in a northern peatland. *Global Change Biology* 13: 397-411.

- , Moore, T., Bubier, J. & Lafleur, P. 1992. Northern fens - Methane flux and climatic change. *Tellus* 44B: 100-105.
- Ruuhijärvi, R. 1983. The Finnish mire types and their regional distribution. In: Gore, A.J.P. (ed.). *Ecosystems of the world 4B mires: Swamp, bog, fen and moor. Regional studies*. Elsevier, Amsterdam. p. 47-68.
- Rydin, H. 1985. Effect of water level on desiccation of *Sphagnum* in relation to surrounding *Sphagna*. *Oikos* 45: 374-379.
- & McDonald, A. J. S. 1985a. Photosynthesis in *Sphagnum* at different water contents. *Journal of Bryology* 13: 579-584.
- & McDonald, A. J. S. 1985b. Tolerance of *Sphagnum* to water level. *Journal of Bryology* 13: 571-578.
- Saarinén, T. 1996. Biomass and production of two vascular plants in a boreal mesotrophic fen. *Canadian Journal of Botany* 74: 934-938.
- Sagerfors, J., Lindroth, A., Grelle, A., Klemetsson, L., Weslien, P. & Nilsson, M. 2008. Annual CO₂ exchange between a nutrient-poor, minerotrophic, boreal mire and the atmosphere. *Journal of Geophysical Research – Biogeosciences* 113(G1): G01001. doi:10.1029/2006JG000306.
- Saveyn, A., Steppe, K., McGuire, M. A., Lemeur, R. & Teskey, R. O. 2008. Stem respiration and carbon dioxide efflux of young *Populus deltoides* trees in relation to temperature and xylem carbon dioxide concentration. *Oecologia* 154: 637-649.
- Schimel, J. P. 1995. Plant transport and methane production as controls on methane flux from arctic wet meadow tundra. *Biogeochemistry* 28: 183-200.
- Schipperges, B. & Rydin, H. 1998. Response of photosynthesis of *Sphagnum* species from contrasting microhabitats to tissue water content and repeated desiccation. *New Phytologist* 140: 677-684.
- Schmid, H. P. 2002. Footprint modeling for vegetation atmosphere exchange studies: a review and perspective. *Agricultural and Forest Meteorology* 113: 159-183.
- Sebacher, D. I., Harriss, R. C. & Bartlett, K. B. 1985. Methane emissions to the atmosphere through aquatic plants. *Journal of Environmental Quality* 14: 40-46.
- Shannon, R. D. & White, J. R. 1994. A three-year study of controls on methane emissions from two Michigan peatlands. *Biogeochemistry* 27: 35-60.
- , White, J. R., Lawson, J. E. & Gilmour, B. S. 1996. Methane efflux from emergent vegetation in peatlands. *Journal of Ecology* 84: 239-246.
- Shevtsova, A., Ojala, A., Neuvonen, S., Vieno, M. & Haukioja, E. 1995. Growth and reproduction of dwarf shrubs in a sub-arctic plant community - Annual variation and aboveground interactions with neighbors. *Journal of Ecology* 83: 263-275.
- Silvola, J. & Aaltonen, H. 1984. Water content and photosynthesis in the peat mosses *Sphagnum fuscum* and *Sphagnum angustifolium*. *Annales Botanici Fennici* 21: 1-6.
- Sjörs, H. 1983. Mires of Sweden. In: Gore, A.J.P. (ed.). *Ecosystems of the world 4B mires: Swamp, bog, fen and moor. Regional studies*. Elsevier, Amsterdam. p. 69-94.
- 1991. Phytomass and necromass above and below ground in a fen. *Holarctic Ecology* 14: 208-218.
- Soegaard, H. & Nordstroem, C. 1999. Carbon dioxide exchange in a high-arctic fen estimated by eddy covariance measurements and modelling. *Global Change Biology* 5: 547-562.
- Sommerfeld, R. A., Mosier, A. R. & Musselman, R. C. 1993. CO₂, CH₄ and N₂O flux through a Wyoming snowpack and implications for global budgets. *Nature* 361: 140-142.

- Strack, M. & Waddington, J. M. 2007. Response of peatland carbon dioxide and methane fluxes to a water table drawdown experiment. *Global Biogeochemical Cycles* 21: GB1007. doi:10.1029/2006GB002715.
- , Waddington, J. M., Rochefort, L. & Tuittila, E. S. 2006a. Response of vegetation and net ecosystem carbon dioxide exchange at different peatland microforms following water table drawdown. *Journal of Geophysical Research – Biogeosciences* 111(G2): G02006. doi:10.1029/2005JG000145.
- , Waddington, J. M. & Tuittila, E. S. 2004. Effect of water table drawdown on northern peatland methane dynamics: Implications for climate change. *Global Biogeochemical Cycles* 18: GB4003. doi:10.1029/2003GB002209.
- , Waller, M. F. & Waddington, J. M. 2006b. Sedge succession and peatland methane dynamics: A potential feedback to climate change. *Ecosystems* 9: 278-287.
- Suyker, A. E., Verma, S. B., Clement, R. J. & Billesbach, D. P. 1996. Methane flux in a boreal fen: Season-long measurement by eddy correlation. *Journal of Geophysical Research – Atmospheres* 101(D22): 28637-28647.
- Svensson, B. H. 1984. Different temperature optima for methane formation when enrichment from acid peat are supplemented with acetate or hydrogen. *Applied and Environmental Microbiology* 48: 389-394.
- , B. H. & Rosswall, T. 1984. In situ methane production from acid peat in plant communities with different moisture regimes in a subarctic mire. *Oikos* 43: 341-350.
- Szumigalski, A. R. & Bayley, S. E. 1996. Decomposition along a bog to rich fen gradient is central Alberta, Canada. *Canadian Journal of Botany* 74: 573-581.
- Tang, J., Baldocchi, D. & Xu, L. 2005. Tree photosynthesis modulates soil respiration on a diurnal time scale. *Global Change Biology* 11: 1298-1304, doi: 10.1111/j.1365-2486.2005.00978.x.
- Tokida, T., Miyazaki, T. & Mizoguchi, M. 2005. Ebullition of methane from peat with falling atmospheric pressure. *Geophysical Research Letters* 32: L13823. doi:10.1029/2005GL022949.
- Treat, C. C., Bubier, J. L., Varner, R. K. & Crill, P. M. 2007. Timescale dependence of environmental and plant-mediated controls on CH₄ flux in a temperate fen. *Journal of Geophysical Research – Biogeosciences* 112(G1): G01014. doi:10.1029/2006JG000210.
- Tuittila, E. S., Vasander, H. & Laine, J. 2004. Sensitivity of C sequestration in reintroduced *Sphagnum* to water-level variation in a cutaway peatland. *Restoration Ecology* 12: 483-493.
- Turunen, J., Tomppo, E., Tolonen, K. & Reinikainen, A. 2002. Estimating carbon accumulation rates of undrained mires in Finland - application to boreal and subarctic regions. *Holocene* 12: 69-80.
- Van Der Nat, F. F. W. A., Middelburg, J. J., Van Meteren, D. & Wielemakers, A. 1998. Diel methane emission patterns from *Scirpus lacustris* and *Phragmites australis*. *Biogeochemistry* 41: 1-22.
- Visser, E. J. W., Bogemann, G. M., Van de Steeg, H. M., Pierik, R. & Blom, C. W. P. M. 2000. Flooding tolerance of *Carex* species in relation to field distribution and aerenchyma formation. *New Phytologist* 148: 93-103.
- Vitt, D. H. & Slack, N. G. 1984. Niche diversification of *Sphagnum* relative to environmental factors in Northern Minnesota peatlands. *Canadian Journal of Botany* 62: 1409-1430.
- Waddington, J. M. & Roulet, N. T. 2000. Carbon balance of a boreal patterned peatland. *Global Change Biology* 6: 87-97.

- , Roulet, N. T. & Swanson, R. V. 1996. Water table control of CH₄ emission enhancement by vascular plants in boreal peatlands. *Journal of Geophysical Research – Atmospheres* 101(D17): 22775-22785.
- Wan, S. & Luo, Y. 2003. Substrate regulation of soil respiration in a tallgrass prairie: Results of a clipping and shading experiment. *Global Biogeochemical Cycles* 17:1054. doi:10.1029/2002GB001971.
- Wang, Z.-P., Han, X.-G., Wang, G. G., Song, Y. & Gullledge J. 2008. Aerobic methane emission from plants in the inner mongolia steppe. *Environmental Science & Technology* 42: 62-68
- Weltzin, J. F., Bridgham, S. D., Pastor, J., Chen, J. Q. & Harth, C. 2003. Potential effects of warming and drying on peatland plant community composition. *Global Change Biology* 9: 141-151.
- Whalen, S. C. 2005. Biogeochemistry of methane exchange between natural wetlands and the atmosphere. *Environmental Engineering Science* 22: 73-94.
- & Reeburgh, W. S. 2000. Methane oxidation, production, and emission at contrasting sites in a boreal bog. *Geomicrobiology Journal* 17: 237-251.
- Wheeler, B. D. & Proctor, M. C. F. 2000. Ecological gradients, subdivisions and terminology of north-west European mires. *Journal of Ecology* 88: 187-203.
- Whiting, G. J. & Chanton, J. P. 1992. Plant-dependent CH₄ emissions in a sub arctic Canadian Fen. *Global Biogeochemical Cycles* 6:225-231.
- & Chanton, J. P. 1993. Primary production control of methane emission from wetlands. *Nature* 364: 794-795.
- & Chanton, J. P. 2001. Greenhouse carbon balance of wetlands: methane emission versus carbon sequestration. *Tellus Series B-Chemical and Physical Meteorology* 53: 521-528.
- Wijesinghe, D. K., John, E. A. & Hutchings, M. J. 2005. Does pattern of soil resource heterogeneity determine plant community structure? An experimental investigation. *Journal of Ecology* 93: 99-112.
- Wilson, D., Alm, J., Riutta, T., Laine, J. & Byrne, K. A. 2007. A high resolution green area index for modelling the seasonal dynamics of CO₂ exchange in peatland vascular plant communities. *Plant Ecology* 190: 37-51.
- Yavitt, J. B. & Lang, G. E. 1990. Methane production in contrasting wetland sites - Response to organic-chemical components of peat and to sulfate reduction. *Geomicrobiology Journal* 8: 27-46.
- , Williams, C. J. & Wieder, R. K. 2000. Controls on microbial production of methane and carbon dioxide in three Sphagnum-dominated peatland ecosystems as revealed by a reciprocal field peat transplant experiment. *Geomicrobiology Journal* 17: 61-88.
- Yurova, A., Wolf, A., Sagerfors, J. & Nilsson, M. 2007. Variations in net ecosystem exchange of carbon dioxide in a boreal mire: Modeling mechanisms linked to water table position. *Journal of Geophysical Research –Biogeosciences* 112(G2): G02025. doi:10.1029/2006JG000342.
- Zinder, S. H. 1993. Physiological ecology of methanogens. In: Ferry, J. G. (ed.). *Methanogenesis: ecology, physiology, biochemistry and genetics*. Chapman & Hall, New York, N.Y. p. 128-206.